Imidazolidin-4-one Derivatives of Primaquine as Novel Transmission-Blocking Antimalarials

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Imidazolidin-4-one derivatives of primaquine were synthesized as potential double prodrugs of the parent drug. The title compounds inhibit the development of the sporogonic cycle of Plasmodium berghei, affecting the appearance of oocysts in the midguts of the mosquitoes. The imidazolidin-4-ones are very stable, both in human plasma and in pH 7.4 buffer, indicating that they are active per se. Thus, imidazolidin-4-ones derived from 8-aminoquinolines represent a new entry in antimalarial structure–activity relationships.

Introduction

Malaria is the major life-threatening parasitic disease in tropical and sub-tropical regions. Worldwide, there are at least 300 million acute cases of malaria and more than 1–2 million deaths each year, mostly young children infected with Plasmodium falciparum.¹ Most of the drugs actually used in antimalarial chemotherapy are particularly active against the asexual blood forms of the parasite, which are responsible for the clinical symptoms of the disease. With the rapid spread of drug-resistant P. falciparum strains, the development of safe and effective antimalarials that prevent transmission, in addition to curing patients, has become an important strategy toward achieving an effective control of malaria.²

In contrast to the asexual blood forms of Plasmodium, the sexual form of the parasite is a much unexplored life-cycle target. Currently, primaquine, ¹, is the only available transmission-blocking antimalarial displaying a marked activity against gametocytes from all species of parasite causing human malaria, including chloroquine-resistant P. falciparum.³ However, the use of primaquine is limited by its extensive conversion to carboxyprimaquine, ², and by its toxic effects, among them hemolytic anemia, particularly in patients who are deficient in glucose-6-phosphate dehydrogenase.⁴–⁶ Several peptide and amino acid derivatives of primaquine and other 8-aminooquinoline antimalarials have been synthesized to reduce the metabolic oxidative deamination pathway, as well as to reduce toxicity of the parent drug.⁷–¹⁰ Such derivatives display improved activity/toxicity ratios when compared to primaquine, which can be ascribed either to a reduction in metabolic inactivation⁸,⁹ or to a selective hydrolysis inside the parasite,⁷ leading to a higher intracellular drug concentration. However, we, and others, have shown that amino acid and peptide derivatives of primaquine are rapidly hydrolyzed to primaquine by aminopeptidases and endopeptidases,⁹,¹¹ suggesting that they might undergo extensive hydrolysis to the parent drug in the GI tract when given orally. One approach to enhance the enzymatic stability of amino acid or peptide derivatives of primaquine toward proteolytic degradation at the mucosal absorption barrier or in the blood is the development of a double prodrug.¹² To this end, imidazolidin-4-one formation was introduced as a useful prodrug approach to protect the N-terminal amino acid residue of di- to pentapeptides against aminopeptidase-catalyzed hydrolysis.¹³–¹⁶ Usually, peptide imidazolidin-4-one derivatives are quantitatively hydrolyzed to the parent peptide in pH 7.4 buffer at 37 °C with half-lives ranging from 1 to 30 h, depending on the N-terminal dipeptide sequence and on the imidazolidinone substituents.¹³–¹⁶ Therefore, we reasoned that imidazolidin-4-one derivatives, ₄, of primaquine (see Scheme 1) would release the corresponding amino acid derivative via a nonenzymatic reaction, which, in turn, could be enzymatically hydrolyzed to primaquine. In this study, we report the reactivity in human plasma and gametocytocidal activity of imidazolidin-4-ones, ₄.

Results and Discussion

Chemistry. The synthesis and characterization of imidazolidin-4-ones ₄ has been described elsewhere.¹⁷ In short, compounds ₄ can be synthesized in good yields from the corresponding amino acid derivatives AA-PQ,
3, by refluxing with an excess of the appropriate ketone in methanol in the presence of triethylamine (TEA) and 4 Å molecular sieves.

**In Vitro Stability Studies.** The hydrolyses of imidazolidin-4-ones 4 in 80% human plasma were monitored by HPLC for the simultaneous disappearance of substrate and formation of the amino acid derivative 3 and primaquine. With the exception of compound 4b, imidazolidin-4-ones 4 display unusually high stability when incubated in 80% human plasma (Table 1), with no significant disappearance of the starting material over 3 days of incubation. The stability of 4 in human plasma is not significantly affected either by the nature of the amino acid R1 substituent or by the R2 and R3 substituents in the imidazolidin-4-one moiety. This contrasts with the behavior of the corresponding amino acid intermediates, 3, which are hydrolyzed quantitatively to primaquine with rates depending on the nature of the amino acid side chain (Figure 1: 4c versus Ala-PQ).

In contrast to their behavior in plasma, the imidazolidin-4-ones 4 hydrolyze to the corresponding amino acid derivatives in pH 7.4 buffer with half-lives ranging from 9 to 30 days (Table 1). Moreover, compounds 4 are hydrolyzed 50–100 times slower than the imidazolidin-4-one counterparts derived from dipeptides or pentapeptides.13–16 We were surprised by such large differences in reactivity, though one possible explanation might lie in the mechanism of hydrolysis. The pH-independent hydrolysis of imidazolidin-4-ones (i.e. at pH 7.4 in 80% human plasma) was monitored (Figure 1).

![Scheme 1](Image)

**Table 1. Percentage of Hydrolysis of Imidazolidin-4-ones 4 to the Corresponding Amino Acid Products 3, in 80% Human Plasma and in pH 7.4 Phosphate Buffer at 37 °C after 3 Days of Incubation, with Half-Lives for the Hydrolysis in pH 7.4 Buffer in Parentheses.**

<table>
<thead>
<tr>
<th>compd</th>
<th>R1</th>
<th>R2</th>
<th>R3</th>
<th>% hydrolysis to 3 after 3 days</th>
<th>80% human plasma</th>
<th>pH 7.4 buffer (t1/2/d)</th>
</tr>
</thead>
<tbody>
<tr>
<td>4a</td>
<td>H</td>
<td>(CH2)5</td>
<td>Me</td>
<td>stablea</td>
<td>11 (18)4b</td>
<td>19 (9.8)</td>
</tr>
<tr>
<td>4b</td>
<td>H</td>
<td>(CH2)5</td>
<td>Me</td>
<td>stablea</td>
<td>16 (12)6c</td>
<td>10 (20)4d</td>
</tr>
<tr>
<td>4c</td>
<td>CH2Ph</td>
<td>(CH2)5</td>
<td>Me</td>
<td>stablea</td>
<td>28 (6.4)6c</td>
<td>17 (11)4d</td>
</tr>
<tr>
<td>4d</td>
<td>CH2Ph</td>
<td>(CH2)5</td>
<td>Me</td>
<td>stablea</td>
<td>6 (31)6f</td>
<td>10 (20)4d</td>
</tr>
<tr>
<td>4e</td>
<td>CH2Ph</td>
<td>Me</td>
<td>Me</td>
<td>stablea</td>
<td>16 (12)6c</td>
<td>6 (31)6f</td>
</tr>
<tr>
<td>4f</td>
<td>CHMe2</td>
<td>(CH2)5</td>
<td>Me</td>
<td>stablea</td>
<td>21 (8.8)6c</td>
<td>8 (26)6f</td>
</tr>
<tr>
<td>4g</td>
<td>CHMe2</td>
<td>Me</td>
<td>Me</td>
<td>stablea</td>
<td>10 (20)4d</td>
<td>8 (26)6f</td>
</tr>
<tr>
<td>4h</td>
<td>CH2CHMe2</td>
<td>Me</td>
<td>Me</td>
<td>stablea</td>
<td>16 (12)6c</td>
<td>8 (26)6f</td>
</tr>
<tr>
<td>4i</td>
<td>CH2CHMe2</td>
<td>Me</td>
<td>Me</td>
<td>stablea</td>
<td>16 (12)6c</td>
<td>8 (26)6f</td>
</tr>
<tr>
<td>4j</td>
<td>CH2CHMe2</td>
<td>Me</td>
<td>Me</td>
<td>stablea</td>
<td>16 (12)6c</td>
<td>8 (26)6f</td>
</tr>
<tr>
<td>4k</td>
<td>CH2CHMe2</td>
<td>Me</td>
<td>Me</td>
<td>stablea</td>
<td>16 (12)6c</td>
<td>8 (26)6f</td>
</tr>
<tr>
<td>4l</td>
<td>CH2CHMe2</td>
<td>Me</td>
<td>Me</td>
<td>stablea</td>
<td>16 (12)6c</td>
<td>8 (26)6f</td>
</tr>
</tbody>
</table>

- No degradation after 3 days of incubation.
- Percentage of hydrolysis after 3 days of incubation.
- From ref 17.
- This study.
- Not determined due to solubility problems.

Figure 1. Time courses for compounds 4c (○), Ala-PQ (■), and primaquine (•) when 4c (open symbols) and Ala-PQ (closed symbols) were incubated in 80% human plasma at 37 °C.

> 4) occurs by an SN1-type mechanism which involves the departure of an amide leaving group (Scheme 2).13 The amides resulting from the rate-limiting ring opening of 4 are much poorer leaving groups than those from dipeptide imidazolidin-4-ones (the difference in pKa is about 3.3 units18). Assuming that the pH-independent hydrolysis of imidazolidin-4-ones has the same susceptibility to the leaving group effect as the analogous acyclic N-Mannich bases,19 i.e. with a Brønsted βlg value of ca. –1, then it would be expected that compounds 4 would hydrolyze ca. 103 times slower20 than their counterparts derived from dipeptides. The smaller differences reported herein might be attributed to the fact that the amino acid chain affects both the amide leaving group ability and the ability of the imidazolidin-4-one N1 amino nitrogen atom to expel the amide. A remark should be made on the higher stability of 4 in human plasma when compared to pH 7.4 phosphate buffer. This might be ascribed to the binding to plasma proteins. A decrease in reactivity in human plasma when compared to pH 7.4 buffers has also been reported for imidazolidin-4-ones derived from dipeptides and pentapeptides.14,15

**In Vivo Gametocytocidal Studies.** The potential of compounds 4 to prevent the transmission of malaria was studied using a model consisting of BalbC mice infected with *Plasmodium berghei* and *Anopheles stephensi* mosquitoes. The two criteria used to assess the antimalarial activity of each compound were (i) the percentage of mosquitoes with oocysts and (ii) the mean number of oocysts per infected mosquito. These in vivo screening assays are of major support in analyzing the effect of drugs in the sporogonic cycle, since no equivalent in vitro assays exist.21 Although this model cannot specifically attribute the drug effect to either gametocytocidal activity (i.e. affects parasite development by killing gametocytes) or sporontocidal activity (i.e. affects directly the development of oocysts on the stomach wall of the mosquitoes), it can clearly show if a compound is effective at interrupting the transmission of the infection to mosquitoes by interference with the cycle in these insects.22,23

The antimalarial activity data are presented in Table 2, and from these the following observations can be made. First, primaquine (PQ) and the imidazolidin-4-one derivatives 4a–g (i.e. those derived from Gly-PQ, Ala-PQ, and Phe-PQ) completely inhibited the produc-
the production of oocysts at 50 \( \mu \text{mol/kg} \). Third, the mean number of oocysts was also significantly affected by compounds 4a–c and 4j–l at a dose of 10 \( \mu \text{mol/kg} \). Primaquine was similarly effective at this dose level. In contrast, at a dose of 10 \( \mu \text{mol/kg} \), derivatives 4d–g (derived from Phe-PQ) and 4h,i (derived from Val-PQ) did not significantly reduce the oocyst production when compared with the control.

From these results it can be concluded that the imidazolidin-4-ones derived from Gly-PQ, 4a,b, and Ala-PQ, 4c, are the most effective gametocytocidal agents, displaying an antimalarial activity comparable to that of primaquine. The activity of 4a and 4b compares to the activity reported previously for Gly-PQ, thus suggesting that incorporation of the imidazolidin-4-one scaffold does not alter substantially the antimalarial activity. In contrast, imidazolidin-4-ones 4 derived from the more lipophilic amino acids phenylalanine, valine, and leucine were less active when compared to primaquine. A similar observation has been reported for dipptide derivatives of primaquine, AA-Gly-PQ, for which it was found that the most active compound was AA = Gly, while those containing the bulky and lipophilic amino acids (R)-Phe and (S)-Phe were less active. More recently, Vangapandu et al. reported that the attachment of a hydrophobic amino acid to the terminal amino group of primaquine analogues results in decreased blood-schizontocidal antimalarial activity.10

Taken together, these suggest that hydrophobic amino acid side chains have a detrimental effect on the activity of 8-aminooquinoline derivatives against both blood schizonts and gametocytes.

The effect of the substituents at the C-2 position of imidazolidin-4-ones 4 on the gametocytocidal activity is less clear. For example, there is no difference in activity between the imidazolidin-4-ones prepared from Phe-PQ and cycloheptanone, 4d, cyclohexanone, 4e, cyclopentanone, 4f, and acetone, 4g, at 10 or 50 \( \mu \text{mol/kg} \). Similarly, no difference in activity is observed between the Gly-PQ derivatives 4a and 4b. In contrast, the cycloheptanone, cyclopentanone, and acetone derivatives of Val-PQ (4h, 4j, and 4k, respectively) are active at 10 \( \mu \text{mol/kg} \), while their cyclohexanone counterpart 4i is inactive at this dose level. This warrants further studies to disclose the effect of the imidazolidin-4-one C-2 substituents on antimalarial activity.

The imidazolidin-4-one N1 nitrogen atom of 4 is substantially less basic (pK\(_a\) ca. 3)13,24 than that of primaquine (pK\(_a\) 10) or its amino acid or peptide derivatives (pK\(_a\) 8–8.5). Consequently, a major finding that emerges from this study is that the presence of a terminal, strongly basic amino group, as found in primaquine or its amino acid and peptide derivatives, is not a necessary requirement for gametocytocidal activity.

**Conclusion**

The reported imidazolidin-4-ones prepared from amino acid derivatives of primaquine exhibit potent gametocytocidal activity against \( P. \) berghei. In general, those derivatives 4 containing small amino acid chains (Gly and (S)-Ala) are superior to those containing bulky/hydrophobic side chains ((S)-Phe, (S)-Val, and (S)-Leu).
These imidazolidin-4-ones 4 are very stable both in chemical and in enzymatic conditions, suggesting that they are active per se. Thus, the imidazolidin-4-ones 4 can be considered as a novel type of 8-aminooxinoline antimalarial. Recent reports indicate that adequate substitution at the C-2, C-4, and C-5 positions of the quinoline moiety can lead to potent 8-aminooxinoline blood-schizontocidal antimalariais devoid of significant blood toxicity. Therefore, combination of the imidazolidin-4-one scaffold with the appropriately substituted quinoline moiety deserves further attention.

**Experimental Section**

**HPLC Analysis.** High-performance liquid chromatography (HPLC) measurements were carried out using a Waters assembly equipped with a model 600 controlled pump and a model 991 photodiode-array detector. A Rheodyne 7725 injection valve equipped with 20-µL sample loop was used. Acquisition and treatment of data were made by means of NEC for MS-DOS, version 3.30 software. The separation was performed on a Purospher, 250 × 4.0-mm i.d. 5 µm (Merck, Germany) analytical column and a LiChrospher 100 RP-8 5 µm (Merck, Germany) employed as precolumn. The solvent system used was a gradient of sodium acetate buffer (pH 4.75, 0.05 M) (A) and acetonitrile (B); 10−3 M triethylamine was added as a preservative. The solvent for the aqueous mobile phase in order to improve peak shape. The gradient was as follows: 0 min, 50% B; 4.5 min, 50% B; 5.0 min, 10% B; 20 min, 10% B. For the imidazolidin-4-one derivatives of valine a second gradient was developed: 0 min, 40% B; 5.5 min, 40% B; 6.0 min, 10% B; 20 min, 10% B. Elution was performed at a solvent flow rate of 1 mL/min, and a 15 mL/min nitrogen sparging was applied to remove dissolved gases. Chromatographic separation was monitored by UV detection at 265 nm. All analyses were performed at room temperature.

**Hydrolysis in Human Plasma.** The compounds 4 were incubated at 37 °C in human plasma (from heparinized blood of healthy donors) diluted to 80% (v/v) with pH 7.4 isotonic phosphate buffer. At appropriate intervals, aliquots were added to acetonitrile to quench the reaction and precipitate plasma proteins. These samples were centrifuged and the supernatant was analyzed by the HPLC method described above for the presence of substrate and products.

**Hydrolysis in Aqueous Solution.** The rates of hydrolyses of compounds 4 were determined in pH 7.4 phosphate saline buffer, at 37 °C. Usually, a 10 µL aliquot of a 10−2 M stock solution of substrate in acetonitrile was added to 10 mL of the appropriate thermostated buffer solution. At regular intervals, samples of the reaction mixture were analyzed by HPLC. All reactions followed first-order kinetics over four half-lives.

**In Vivo Gametocytocidal Activity.** BalbC mice were infected by intraperitoneal inoculations of 106 erythrocytes parasitized with *P. berghei* ANKA. After 4 days, when the presence of gametocytes and exflagellation was observed by microscopic observation of Giemsa stained blood films, mice were randomly separated into five different groups of six animals. Each group was treated by intraperitoneal administration with one single dose of each compound 4 and primaquine (10 and 50 µmol/kg in inoculation volumes of 0.1–0.2 mL; controls consisted of mice given a PBS solution). Two hours after administration, mice were anesthetized and placed on top of individual cages containing ca. 50 glucose-starved *Anopheles stephensi* female mosquitoes, which were allowed to feed for 2 h. After the blood meal, uninfected females mosquitoes were removed from each cage. Ten days after the blood meal, 10 µL aliquots of each cage were randomly collected and dissected for microscopic detection of oocysts in midguts. For further details see ref 9.

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(20) Assuming that the equation \( \log k = -pK_a + C \) holds for compounds 4, then \( \log(k_1/k_2) = pK_a^2 - pK_a^1 \).


(24) For example, the \( pK_a \) value for the derivative 4f, kinetically determined from a pH–rate profile, is 3.5 (data not shown).