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MESTRADO INTEGRADO EM MEDICINA

Metabolomic signatures after bariatric surgery – a systematic review

Sara Matilde Branco Vaz



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Sara Matilde Branco Vaz

sarambvaz@gmail.com

Mestrado Integrado em Medicina

Instituto de Ciências Biomédicas Abel Salazar – Universidade do Porto

Orientadora

Professora Doutora Mariana P. Monteiro

Professora Associada

Instituto de Ciências Biomédicas Abel Salazar – Universidade do Porto

Coorientadora

Professora Doutora Sofia S. Pereira

Professora Auxiliar

Instituto de Ciências Biomédicas Abel Salazar – Universidade do Porto

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Sava Matilde Bancolaz Sara Matilde Branco Vaz

in مريئة

Mariana P. Monteiro

) in Tereira Sofia S. Pereira

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Resumo

<u>Introdução</u>: A metabolómica tem vindo a surgir como uma ferramenta poderosa, ao possibilitar a compreensão das respostas do organismo a diferentes intervenções terapêuticas. A cirurgia bariátrica é o tratamento mais eficaz para a obesidade e respetivas comorbilidades.

<u>Objetivo</u>: O nosso propósito foi a elaboração de uma revisão sistemática tendo por base os dados disponíveis relativamente aos perfis metabolómicos que caracterizam os pacientes submetidos a diferentes tipos de cirurgia bariátrica. Como objetivo adicional foram revistos os perfis metabolómicos com utilidade na previsão da remissão da diabetes tipo 2 e das diferenças na perda de peso após a cirurgia.

<u>Metodologia</u>: Foram seguidas as guidelines PRISMA - Preferred Reporting Items for Systematic Reviews and Meta-Analyses - para a execução da revisão sistemática. Quarenta e seis estudos satisfizeram os critérios de elegibilidade. Foram compilados e resumidos os dados de artigos originais, cujo conteúdo se focava nos perfis metabolómicos induzidos pela cirurgia bariátrica.

<u>Resultados</u>: Os metabolitos mais frequentemente influenciados pela cirurgia bariátrica são os aminoácidos, lípidos, metabolitos relacionados com o metabolismo energético e metabolitos associados à microbiota intestinal. Dentro destes, os padrões pré-operatórios e pós-operatórios imediatos dos aminoácidos de cadeia ramificada, aminoácidos aromáticos, metabolitos do ciclo do ácido tricarboxílico, fosfolípidos, ácidos gordos de cadeia longa e ácidos biliares foram aqueles que tiveram uma associação mais significativa com as diferenças nas respostas induzidas após a cirurgia.

<u>Conclusões</u>: O perfil metabolómico pode vir a destacar-se como um forte auxiliar para a previsão de respostas a longo prazo, consequentes dos diferentes tipos de cirurgia bariátrica. Deste modo, poderá possibilitar uma maior personalização nos tratamentos, levando a uma otimização na alocação dos recursos para a saúde.

<u>Palavras-chave:</u> Metabolómica · Cirurgia Bariátrica · Resultados terapêuticos · Perda de peso · Diabetes tipo 2

Abstract

<u>Introduction</u>: Metabolomics emerged as an important tool to gain insights on how the body responds to therapeutic interventions. Bariatric surgery is the most effective treatment for severe obesity and obesity-related co-morbidities.

<u>Purpose</u>: Our aim was to conduct a systematic review of the available data on metabolomics profiles that characterize patients' submitted to different bariatric surgery procedures, which could be useful to predict type 2 diabetes remission and weight loss and outcomes.

<u>Methods</u>: The Preferred Reporting Items for Systematic Reviews and Meta-Analyses - PRISMA guidelines were followed. Forty-six studies successfully met the eligibility criteria. Data from original study reports addressing metabolomics profiles induced by bariatric surgery were compiled and summarized.

<u>Results:</u> Amino acids, lipids, energy metabolism-related metabolites and gut microbiota-related metabolites were the metabolite classes that were most frequently influenced by bariatric surgery. Among these, branched chain amino acids, aromatic amino acids, tricarboxylic acid cycle metabolites, phospholipids, long-chain fatty acids and bile acids were the ones which preoperative or early post-operative pattern, seems to better correlate with bariatric surgery outcome.

<u>Conclusions</u>: Metabolite profiling could become a useful tool to predict long term response to different bariatric surgery procedures, allowing more personalized interventions and improving healthcare resources allocation.

Keywords: Metabolomics · Bariatric Surgery · Clinical outcome · Weight loss · Type 2 Diabetes

Abbreviation List | Lista de abreviaturas

3-HAA	3-hydroxyanthranilic acid
5-HIAA	5-hydroxyindoleacetic acid
5-HTrp	5-hydroxytryptophan
AA	Amino Acid
AAA	Aromatic Amino Acid
AC	Acylcarnitine
BA	Bile Acid
BCAA	Branched Chain Amino Acid
BMI	Body Mass Index
BPD	Biliopancreatic Diversion
BPL	Biliopancreatic Limb
CDCA	Chenodeoxycholic Acid
DCA	Deoxycholic Acid
DJB	Duodenal Jejunal Bypass
DS	Duodenal Switch
FFA	Free Fatty Acid
FXR	Farnesoid X Receptor
G-	Glycine amidated sub-fraction
HbA1c	Glycated Hemoglobin
HCA	Hyocholic Acid
HDL	High-density Lipoprotein
HOMA-IR	Homeostasis Model Assessment–Insulin Resistance
L-DOPA	L-dihydroxyphenylalanine
LAGB	Laparoscopic Adjustable Gastric Band
LDL	Low-density Lipoprotein
LPC	Lysophosphatidylcholines
MCA	Muricholic Acid
MCSFA	Medium-Chain Saturated Fatty Acids
NEFA	Non-Esterified Fatty Acids
PC	Phosphatidylcholines
PE	Phosphatidyle than olamines
PL	Phospholipid
QUICKI	Quantitative Insulin Sensitivity Check Index
RYGB	Roux-en-Y Gastric Bypass
SADI-S	Single-anastomosis duodeno-ileal bypass with Sleeve Gastrectomy
SG	Sleeve Gastrectomy
SM	Sphingomyelin
T2D	Type 2 Diabetes Mellitus
TCA	Tricarboxylic Acid

TLR4Toll-Like Receptor 4TMAOTrimethylamine N-OxideVLDLVery-low-density Lipoprotein

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Introduction | Introdução

The increasing prevalence of obesity in modern society makes it one of the main public health concerns¹. Obesity is a major risk factor for several other medical conditions and particularly for non-communicable diseases, which besides having a great negative impact on the individual's quality of life, are also responsible for the majority of deaths in western countries²⁻⁴. Bariatric surgery is the most effective treatment for severe obesity and its associated co-morbidities, since it has proven to successfully achieve a significant and sustained body mass index (BMI) decrease, besides improving several obesity related diseases, such as diabetes, hypertension, dyslipidemia and cardiovascular events risk⁴. Consequently, bariatric surgery not only improves the health status but also reduces overall mortality and increases the life expectancy of patients with obesity.

Metabolomics has arisen as a valuable tool to study the metabolome dynamic adaptations³. Metabolomics has been widely used across several scientific areas in the quest for molecular fingerprints that could act as diagnostic and prognostic biomarkers. Metabolomics analyses are usually performed by stand-alone hydrogen nuclear magnetic resonance technique or mass spectrometry technique, combined with different metabolite chromatographic separation methods, such as capillary electrophoresis, liquid or gas chromatography⁵. These metabolite detection methods allow the characterization of low molecular weight metabolites from several different classes in a qualitative and quantitative approach⁵. The use of metabolomics in the study of obesity has empowered, not only the understanding of the biochemical and metabolic disruptions underlying this disease condition, but also the acknowledgement of the metabolic and physiological impact of therapeutic interventions, such as bariatric surgery⁶. Metabolomics studies enable to evaluate to what extent does the anatomical modifications of the gastrointestinal tract induced by bariatric surgery alter the individual's metabolomics profile⁵. Furthermore, these techniques also allow to understand how different bariatric surgery techniques that result in distinct anatomical rearrangements of the gut, impact on the individuals' metabolic profiles⁷. Some metabolomics signatures harbor the potential to provide a mechanistic explanation on the heterogeneity of patient outcomes elicited by bariatric surgical interventions^{8,} 9.

Thus, the aim of this review was to systematize the available data in order to describe the metabolic fingerprints that characterize patients' submitted to different bariatric surgery procedures. Our goal was also to identify a pre-operative or early postoperative metabolite

profile potentially useful to predict weight loss response and type 2 diabetes (T2D) remission after bariatric surgery.

Methods | Métodos

Protocol and registration

This project was submitted to PROSPERO (registration number CRD42021235341) and it is available at https://www.crd.york.ac.uk/prospero/display_record.php?ID=CRD42021235341. The review was performed according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines¹⁰.

Information sources and search approach

Publications reporting original data on metabolomics profiles induced by bariatric surgery were searched in three different electronic bibliographic databases: PubMed, Scopus and Isi Web of Knowledge, in February 2021. The detailed search approaches for each database are described in the Supplementary File 1.

Study selection and inclusion criteria

The main domain of this review were metabolomics studies carried out in patients submitted to bariatric surgery for the primary treatment of obesity and obesity related-disorders. The inclusion criteria comprised studies conducted in adult individuals submitted to bariatric surgery, including Sleeve Gastrectomy (SG), Laparoscopic Adjustable Gastric Band (LAGB), Roux-en-Y Gastric Bypass (RYGB), Duodenal Jejunal Bypass (DJB), Biliopancreatic Diversion (BPD), Duodenal Switch (DS), Single Anastomosis with DS, and Single-anastomosis Duodeno-ileal Bypass with Sleeve Gastrectomy (SADI-S). The exclusion criteria were study data pertaining to children (under 18 years old) and pregnant women.

Pre-established restrictive conditions for article retrieval and eligibility were: type of experimental design (only original observational studies, including cohort and case control), species submitted to analysis (only human species), type of biological fluid used for metabolomics analysis (plasma or urine) and publication language (English). No publication date restrictions were applied.

<u>Data extraction</u>

Studies provided by the bibliographic databases were screened to eliminate duplicates. Afterwards, the publications were independently reviewed by two authors, based on title and abstracts, in order to assess the eligibility criteria of each study. Thereafter, full text examination was also required in case of doubt. A third author conducted an independent review of the discordant articles and solved disagreements by majority consensus. The papers selected for inclusion were scored according to the Newcastle-Ottawa Scale, for cohort or cross-sectional studies as appropriate, to assess the quality and risk of bias. Only studies considered to be at least of moderate quality, i.e. with a score of 6 or higher, were included for data analysis. The corresponding scores for each final selected study are described in Table 1.

The selected studies were divided among authors for individual data extraction, and later reviewed by other author in a cross-over manner. Data was retrieved and summarized according to the following information: author(s), study name, reference, experimental design, number of subjects enrolled, most relevant patient features (age, gender proportion, BMI and presence of T2D), type of bariatric surgery procedure, type of biological fluid used for metabolomics analysis, time points of the sample collection, experimental approach, study outcomes when applicable and major findings.

The articles were also subdivided into four different groups: 1) studies considering pre-operative and post-operative metabolomic signatures; 2) studies that disclosure pre-operative and early post-operative metabolomic signatures associated with T2D; 3) studies that disclosure preoperative and early post-operative metabolomic signatures associated with weight loss response; 4) studies comparing metabolomic profiles after different bariatric surgery interventions.

Results | Resultados

Our search identified 378 papers in Scopus, 227 in PubMed and 233 in Isi Web of Knowledge, resulting in a total of 838 papers. After elimination of duplicates (n=328), a total of 510 papers were submitted to an initial screen by reading papers' titles and abstracts by two independent researchers. From those, 461 were out of scope resulting in a total of 49 full-text papers to be evaluated for eligibility. After reading the full texts, 6 papers were eliminated due to the following reasons: repeated data (n=1), out of scope (n=3), no description of metabolites analyzed (n=1) and metabolomics studies conducted in biological fluids other than urine or blood (n=1). Additionally, 3 papers were identified from reading the full texts reference list, resulting in a total of 46 papers to be included in the systematic review (Figure 1). Among these, 45 studies included conducted metabolomic analysis on plasma or serum and only 1 study was conducted on urine.

1) Metabolomic profiles induced by bariatric surgery

Several studies focused on evaluating the changes in metabolomic profile induced by bariatric surgeries (Table II). Amino acids, lipids, energy metabolism-related metabolites and gut microbiota-related metabolites were the most frequently studied metabolite classes. For simplification purposes, only metabolites reported to be more consistently altered after bariatric surgery are presented in this section.

Amino acids are one of the metabolite classes that is most affected by bariatric interventions. Numerous studies demonstrated that circulating levels of valine¹¹⁻²⁶, isoleucine^{11-18, 20, 21, 23-28}, leucine^{11-18, 20, 21, 23-29}, phenylalanine^{11, 12, 14, 15, 17, 18, 20-23, 29}, tyrosine^{11-15, 17, 18, 21, 23, 24, 29}, tryptophan^{11, 15, 18, 23, 29, 30}, alanine^{11-15, 17, 22, 24, 31}, proline^{11-13, 15, 22, 31}, methionine^{12, 17, 24}, aspartate¹⁵, threonine^{13, 15, 26}, lysine¹³ and ornitine^{12, 15, 18, 24, 32} decrease after bariatric surgery. Contrarily, circulating levels of glycine^{12-16, 18, 20, 21, 25, 28, 30, 33} and serine^{12, 22, 25, 30} were reported to increase after bariatric procedures. While other AAs, such as glutamine, histidine, arginine and asparagine were inconsistently reported as being either increased ^{14, 18, 21, 28, 33, 34} or decreased ^{12, 13, 15, 17, 22, 24, 31} after bariatric surgery. No pattern, regarding the type of surgical procedures or the post-operative time, was recognized as potentially responsible for the differences in these specific AA profiles observed across the studies.

Among the lipid class, acylcarnitines (ACs), free fatty acids (FFAs), bile acids (BAs) and phospholipids (PLs) were the metabolites most frequently studied in patients submitted to bariatric surgery interventions.

Acylcarnitines are esters of L-carnitine and fatty acids. These molecules can be divided into three different subclasses: short-, medium and long-chain ACs. The short chain AC acetylcarnitine was reported to increase rapidly after interventions and to remain elevated for 6 to 12 months after bariatric surgery^{11, 15, 22, 26, 29, 35}. Contrarily, the short chain acylcarnitines derived from the branched chain amino acids (BCAAs) catabolism, C3 and C5, were shown to decrease after bariatric surgery^{11, 12, 22, 24, 35}. A transient increase in long-chain ACs levels was observed two-weeks after surgery ^{22, 36}, while most studies report a decrease in medium- and long-chain ACs over the long-term in patients submitted to bariatric surgery^{11, 12, 34, 35}.

Analysis of circulating fatty acids profiles identified a decrease in unsaturated and long-chain saturated fatty acids and non-esterified fatty acids (NEFA) after bariatric surgery, despite some inconsistent results across different studies^{11, 13, 16, 19, 26, 37}. The opposite pattern was observed for medium-chain saturated fatty acids (MCSFA), in particular for the decanoic acid, which was found to increase after surgery^{14, 16, 19, 26, 29}.

Fasting primary and secondary circulating BA levels were also found to increase significantly after bariatric surgery interventions with a malabsorption component, namely RYGB and BPD^{36, 38, 39}. This effect was demonstrated to be even more pronounced in the post-prandial for total BA, glycine- and taurine-conjugated BA⁴⁰. In contrast, studies conducted in patients submitted to restrictive procedures yielded inconsistent results, with some studies reporting no differences between pre- and post-operative BA levels and others reporting increased levels in fasting and post-prandial level of glycine conjugated-BA^{39, 41}.

Phospholipids are important molecules within the cell membrane structure, which include several subclasses such as sphingomyelins (SMs), lysophosphatidylcholines (LPCs), phosphatidylcholines (PCs) and phosphatidylethanolamines (Pes). SM is the dominant sphingolipid in mammalians cells membranes. After BPD, most saturated and unsaturated SMs were found to be decreased³⁹. A decrease in saturated SMs levels^{35, 42} and increase in unsaturated SMs levels^{29, 35, 42} after RYGB was identified by the majority of studies, although results are not consistent across reports^{11, 15, 19, 29, 35-37, 42}.

LPC, PC and PE levels were also reported as being overall decreased after bariatric surgeries with a malabsorptive component, RYGB and BPD, by the majority of studies^{11, 19, 29, 36, 37, 42}, although post-operative increase in some species of PLC and PE was also reported ^{11, 13, 39}. Noticeably, most of the aforementioned PLs were found to increase after SG³⁹.

Likewise, an increase in ketone bodies (3-hydroxybutyrate and acetoacetate) levels was also identified after bariatric surgery^{13, 21, 29, 30, 35, 43}. Although, carboxylic acids and ketone bodies levels were only observed or were more pronounced in short-term period after surgery^{16, 21}.

Most studies reported a decrease in pyruvate^{21, 22, 27, 33} and lactate^{13, 22, 26} levels after bariatric surgery. Contrarily, the Krebs cycle intermediates (citrate ^{13, 18, 21, 29}, succinate ^{18, 30}, fumarate ^{30, 33}, malate ^{18, 33}, oxalacetate ²⁷) were found to increase after surgery.

Gut microbiota metabolites related to AAs and carnitine metabolism were also found to increase after bariatric surgery. In particular, in the levels of p-cresol ^{11, 13, 26, 29}, indole and indoxyl sulfate^{14, 19, 23}, trimethylamine N-oxide (TMAO)^{14, 21}, phenol sulfate ²⁹, 3-indolelactic acid ²⁹ and 4-hydroxy-L-proline²⁹.

2) Metabolomic signatures associated with post-bariatric T2D remission

Metabolomics profiles associated with glucose and insulin homeostasis after the bariatric surgery were analyzed in fourteen independent prospective cohort study studies. While seven papers focused on T2D remission after bariatric surgery, another six studies addressed the specific profile associated with T2D related parameters, namely glycated hemoglobin (HbA1c) levels, insulin secretion and insulin sensitivity, as inferred from insulinogenic index, homeostasis model assessment-insulin resistance (HOMA-IR)⁴⁴ and QUICKI.²³ The majority of studies included individuals submitted either to SG^{23, 45, 46} or RYGB^{11, 14, 19, 44, 47-50}. The remaining three articles were conducted in individuals submitted to different bariatric surgery procedures having RYGB as constant comparator^{38, 51, 52}. (Table III) (Figure 2)

Circulating AAs was the most differentially altered metabolite class distinguishing patients that achieved T2D remission from those with persistent disease after bariatric surgical interventions^{23, 45-47, 50, 51}. BCAAs decrease after SG and RYGB, being more pronounced in patients achieving T2D remission^{23, 44, 46, 47, 50}. *Kwon Y et al* described, not only an early decline in BCAAS, but also aromatic amino acids (AAAs)²³ in patients depicting improved insulin resistance 3 months after SG. The same authors also documented higher pre-preoperative levels of kynurenine pathway metabolites, such as 3-HAA, xanthurenic acid, anthranilic acid and 3-hydroxykynurenine, in patients who later experienced a considerable decrease of these metabolites along with improved insulin secretion and resistance²³. Another study highlighted that patients who achieved T2D remission at 1 year after SG and RYGB had higher baseline levels of L-dihydroxyphenylalanine (L-DOPA) and 3-hydroxyanthranilic acid (3-HAA)⁵¹.

Higher pre-operative levels of tryptophan¹¹ and tricarboxylic acid (TCA) cycle metabolites (citric acid, fumarate and aconitate)¹⁹ were reported in patients with better T2D outcomes. Tryptophan-derived gut microbial metabolites after surgery also differed, as decreased indole-3-propionic acid and increased indole-3-acetic acid were shown to correlate with improved insulin homeostasis 3 months after intervention²³.

Pre-operative lipid metabolite fingerprints of patients who achieve T2D remission as compared to those without disease remission was shown to be characterized by higher PEs, triglycerides and PLs with long-chain fatty acids¹⁹; while postoperative metabolite profile was characterized by lower VLDL, LDL, N-acetyl glycoproteins and unsaturated lipids, and increased HDL and PCs ^{11, 45, 52}. *Zhao L et al* described that higher baseline ratios of stearic acid/palmitic acid and eicosadienoic acid/linoleic acid characterized patients undergoing T2D remission 1 year after RYGB. A substantial decrease in long chain fatty acids (FFA 16: 0, FFA 18: 3 and FFA 17: 2) levels at 6 months after RYGB was identified in patients experiencing T2D remission 1 year after surgery. ¹¹ Other authors established that higher levels of 3-hydroxybutyrate 1 week after bariatric surgery (RYGB and DJB) was associated with greater HbA1c improvements 3 months later⁵² and lower 2-hydroxybutyric acid at 6 months after RYGB was associated with greater BYGB was associated with HOMA-IR improvement⁴⁴. *Arora T et al* studied the metabolic profile of patients submitted to RYGB who experienced T2D remission 2 years after surgery. Postoperative fasting plasma lipid profile was characterized by a significant reduction of most lipid species at 4 days and an increase of the same metabolites at 42 days after surgery.

The utility of bile acids levels as biomarkers of T2D improvement after bariatric surgery was explored in three independent studies^{11, 48}. Two studies addressing T2D remission after RYGB identified higher total BA levels at baseline in the sub-groups of patients with better T2D outcomes^{11, 38, 48}. *Yu H et al* even highlighted that Chenodeoxycholic Acid (CDCA) and the ratio of CDCA/total BA correlated with T2D remission 2 years after RYGB. *Ahlin et al* found that increased total BA levels at 185.3 (±72.9) days after RYGB and BPD were correlated insulin resistance improvement regardless the type of surgical procedure³⁸.

3) Metabolomic signatures associated with post-bariatric weight loss response

The analysis of pre- and post-operative metabolomics according to post-bariatric weight loss responses was assessed in 4 independent prospective studies, conducted in patients submitted to SG^{41, 53} or RYGB^{14, 54}. (Table III) (Figure 3)

AAs were the metabolic class that portrait more pronounced changes after bariatric surgeries^{14, 53, 54}. Higher baseline BCAAs levels, in particular isoleucine, were correlated with greater weight

loss at 3 and 6 months after SG⁵³. In the postoperative period, there was a significant decrease in creatine, ornithine, arginine and valine levels in the sub-group of patients with greater weight loss 1 year after RYGH¹⁴.

Moreover, *Abidi W et al* compared metabolite pattern of patients with sustained weight loss with those of patients who experienced weight regain years after RYGB. The study unraveled lower levels of metabolites related to serine, glycine and threonine pathways, from phenylalanine alanine and glutamate metabolism and TCA cycle, as well as higher levels of other AAs, characterized the metabolomics profile of patients with poor long-term weight loss outcomes. While higher glycine levels were found in patients with sustained weight loss⁵⁴.

Serotonin, a molecule derived from the AA tryptophan, and its metabolites were also proposed as weight loss response molecular fingerprints. A pre-operative profile with lower serotonin levels and serotonin/5-HTrp ratio, and higher 5-HIAA levels and 5-HIAA/serotonin ratio was identified in patients with greater weight loss at 3 and 6 months after SG⁵³.

Patients with sustained weigh loss also presented upregulated lipolysis and consequently depletion of triacylglycerols, diaglycerols and cholesterol esters, in addition to higher levels of 3-hydroxybutyrate⁵⁴.

Increase in multiple BAs subtypes, more pronounced in postprandial rather than fasting period, was found to correlate with early (6 weeks) and short-term (12 weeks) weight loss response after SG and persistent. During fasting, HCA (total, unamidated and glycine amidated subtypes) was the only BA found to be increased, while the postprandial profile portrayed increased BAs (total and glycine amidated subtypes-), CDCA (total and glycine amidated), DCA (total and glycine amidated), MCA (total and glycine amidated), HCA (total, unamidated and glycine amidated). The authors highlighted the augmented post-prandial total and G-CDCA, as being significantly correlated to the 6 weeks BMI loss, as well as the increased post-prandial G-HCA, which was significantly correlated to a greater weight loss percentage at both 6 and 12 weeks ⁴¹.

4) Metabolomic profiles induced by different bariatric surgeries

Only reports comparing the metabolomic profiles induced by different types of bariatric surgery procedures from parallel arm studies were included in this review section. Nine prospective cohort studies and two cross-sectional studies compared the metabolomic profiles induced by different bariatric procedures ^{12, 21, 25, 29, 31, 38, 39, 46, 55} or its variants^{21, 56}. (Table IV) From those, seven studies compared the effects of RYGB with two different restrictive bariatric surgeries: LAGB^{12, 25, 42} and SG^{21, 29, 31, 46}, at several time points after surgery, ranging from 3 days to 1 year.

Metabolomic signatures associated with each type of bariatric surgery procedure were found. However, one study reported that metabolic signatures differences between RYGB and SG tend to be less prominent 12 months after surgery²⁹. When comparing the two types of surgeries, changes in AAs and gut microbiota-related metabolites were the most differentially altered metabolite classes^{21, 25, 29, 31, 46}. In addition, a rapid decrease in the majority of lipid classes was observed after both RYGB and LAGB surgeries in the short-term (1 month). However, some PC and SM species returned or tended to return to baseline values 3 months after LAGB, but not after RYGB⁴².

One study that compared the effect of restrictive (SG) and malabsorptive (BPD) bariatric surgeries on metabolomics found that sphingolipids, PLs and BAs levels were differentially altered by the two bariatric surgery procedures. BPD induced an overall decline in sphingolipids and PLs and an increase in BAs levels. Contrarily, SG induced an increase in sphingolipids and PLs and no changes in BAs levels³⁹.

In contrast, in another study comparing RYGB and BPD, BAs levels were found to be similar after both surgeries³⁸.

Comparison of fasting and post-prandial metabolomics profile of patients submitted to two different malabsorptive surgeries (SADI-S and BPD-DS) was explored in a cross-sectional study. Higher post-prandial BCAAs levels after SADI-S, the least malabsorptive surgery, was the only difference observed⁵⁵.

The effects of two different RYGB variants on circulating metabolomics profiles was evaluated in a cross-sectional study and in another prospective study. However, the differences in RYGB limb lengths used in the two studies do not allow direct comparisons⁵⁶. *Gralka et al* included patients submitted either to a proximal RYGB consisting of a biliopancreatic limb (BPL) of 60 cm and an alimentary limb of 150cm; or to a distal RYGB that had both BPL and common limb lengths of 60-100 cm^{21, 56}. In contrast, the RYGB variants described by *Jarak et al* had different BPL length (short-BPL: 60-100cm vs long-BPL: 200cm) but the same alimentary limb length (120 cm)⁵⁶. Despite the dissimilarities between RYGB variants and no differences on the post-operative AAs levels were noticed, both studies found diverse changes in gut microbiota-related metabolites, namely acetate and dimethyl sulfate, when comparing the two RYGB variants^{21, 56}.

Discussion | Discussão

High levels of circulating AAs is a well-known feature of patients with obesity⁵⁷. BCAAs levels are particularly high and experience a rapid decrease after bariatric surgery⁵⁸ ¹¹⁻²⁹. The reduction in BCAA has been attributed to a combination of multiple factors including decreased protein intake; decreased in amino acids absorption; increased BCAA catabolism and decreased protein catabolism as a consequence of insulin sensitivity improvement and metabolic amelioration³. The upregulation of BCAAs catabolism is supported by the consistent postoperative decline of specific subproducts of BCAAs' mitochondrial oxidation, namely short chain acylcarnitine's C3 and C5.^{11, 12, 22, 24, 35} Although BCAAs decrease after both restrictive and malabsorptive surgeries ¹¹⁻²⁹, parallel arm studies reported a greater effect of RYGB in BCAAs levels.^{21, 25} Since the differences found in BCAA levels when comparing RYGB and the restrictive procedures seem to be independent of the weight loss, these were hypothesized to be related to impaired AAs absorption induced by the RYGB intestinal rearrangement^{21, 25}.

Similarly, AAAs were also found to be elevated in patients with obesity and decrease after bariatric surgery^{21, 29}. This is consistent with the increase of AAAs derived gut microbiota metabolites (p-cresol, indoxyl sulfate, phenol sulfate and 3-indolelactic acid) after bariatric surgery^{11, 13, 14, 23, 26, 29}.

Circulating essential AAs levels were also associated with weight loss response after bariatric surgery, given that higher AAs levels were reported in patients experiencing weight regain⁵⁴. On other hand, patients with significant weight loss presented lower pre-operative levels of serotonin, suggesting its potential use as prognostic biomarker to identify patients that are more likely to achieve greater weight loss after surgery⁵³. Circulating serotonin interacts with multiple organs, preparing the body for energy storage by promoting insulin secretion and de novo lipogenesis in the liver and white adipose tissue, while reducing lipolysis⁵⁹.

Additionally, multiple studies reported that BCAAs and AAAs modifications induced by the bariatric procedures could play a relevant role predicting T2D remission^{23, 45-47, 51}. Obesity associated hyperaminoacidemia is a consequence of insulin resistance, which positively affects protein synthesis and proteolysis⁶⁰. BCAAs can also modulate insulin secretion and promote diabetes via hyperinsulinemia, considering its role as insulin secretagogues. Chronic hyperinsulinemia can further stimulate compensatory insulin resistance and potentially lead to pancreatic β -cell exhaustion^{61, 62}. Contrariwise, improved insulin sensitivity and secretion after bariatric surgery are associated with decreased circulating AAs^{20, 23, 47.} However, relevant pre-

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operative metabolomic profiles are those that attract the greatest interest in order to identify patients that are more likely to benefit from bariatric treatment interventions. Hence, baseline levels of L-DOPA, 3-HAA, tryptophan and TCA cycle subproducts are the metabolites that demonstrated a potential prognostic value for T2D remission^{11, 19, 51}.

In opposition to most of AAs, the levels of the non-essential AA glycine and serine are known to be low in subjects with obesity and increase after bariatric surgery^{12-16, 18, 20-22, 25, 28, 30, 33}. In healthy subjects, glycine is inter-convertible with serine. Glycine participates in multiple biological functions, such as the glutathione synthesis, purines and primary bile salts⁶³. Previous studies described low fasting levels of glycine in individuals with impaired glucose tolerance and proposed glycine as an early marker for insulin resistance⁶⁴. Increased levels of glycine after bariatric surgery suggests a reduction in insulin resistance and oxidative stress¹⁶. In addition, glycine and serine levels were noticeably significantly higher in patients with sustained weight loss, as compared to patients with weight regain⁵⁴. In further support of the potential role of glycine in mediating weight loss, a recent animal model study also concluded that glycine supplementation to obese mice during calorie restriction accelerated fat loss and protected against muscle loss⁶⁵.

Gut microbiota plays an important role in various physiological processes, including metabolism of dietary components and some host-generated substances, with an impact on the use and storage of energy⁶⁶. Gut microbiota of individuals with obesity is characterized by a reduced bacterial diversity and lower ratios of Bacteroidetes to Firmicutes⁶⁷. After bariatric surgery, different gut microbiome related metabolites arise that differ depending on the type of bariatric procedure. In particular, p-cresol, a metabolite of phenylalanine and tyrosine fermentation by gut microbiota^{68, 69}, was reported to increase after SG²⁹. Bacteroides fragilis is one of the bacteria responsible for phenylalanine and tyrosine fermentation. Bacteroides were previously demonstrated to increase after SG in individuals with prior T2D diagnosis⁷⁰. So, the effect in tyrosine and phenylalanine levels found of patients with T2D after SG when compared to RYGB, may be related to increased fermentation by gut microbiota. Sulfate containing metabolites and TMAO, two other gut microbiome related metabolites were identified to increase after RYGB but not after SG²⁹. The differences observed in sulfate containing metabolites results could easily be justified by the fact that the major group of sulfate-reducing bacteria is mainly present in the duodenum, which is trespassed in RYGB⁷¹. Contrary, the effects of RYGB in TMAO levels is not well understood and unexpected because high TMAO levels have been linked to cardiovascular diseases, whereas RYGB is known to reduce the cardiovascular risk and events^{72, 73}. The increase in TMAO levels observed after RYGB, but not after SG, could be derived from the shortening of the small bowel and a less anaerobic metabolism by the gut microbiota, in result of the increase in microbes, such as E. coli and Pseudomonas, responsible for raising TMAO levels⁷⁴. Gut microbiota-related metabolites were also differentially altered by RYGBs with different absorptive limbs. *Gralka et al* found that RYGB with a shorter common limb led to a higher increase in dimethyl sulfate levels compared to a more absorptive RYGB²¹. A different study also found that post-prandial levels of acetate were higher in patients submitted to RYGB with a longer BPL⁵⁶. Both metabolites (dimethyl sulfate and acetate) are produced in the colon by bacterial fermentation⁷⁵, which is enhanced by the higher quantity of undigested food that reaches the colon in the less absorptive RYGB.

The obesity associated energy metabolic disruption is characterized by decreased oxidation of ketone bodies and down-regulation of TCA cycle⁷⁶. Metabolomic analysis suggests that after bariatric surgery a strong and early upregulation of catabolism and lipolytic activity occurs. Most of intermediate and end products of β -oxidation and ketogenesis increase after intervention, in particular in short-term period after surgery^{13, 21, 22, 29, 30, 35, 36, 43}. These findings support a shift in energy metabolism from anabolic to catabolic status, possibly triggered by surgery and caloric restriction.

The increased levels of ketone bodies after bariatric surgery are consistent with the rapid decrease in ketogenic amino acids after bariatric surgery. The ketone body 3-hydroxybutyrate originates from fatty acids degradation for energy generation in the liver⁷⁷. It has been previously reported that 3-hydroxybutyrate levels increase during the first week after bariatric surgery, as a consequence of the acute and adaptative fasting-like response¹⁶. Therefore, individuals with sustained long term weight loss after surgery maintain higher metabolic needs and fatty acid oxidation, responsible for the higher levels of 3-hydroxybutyrate observed⁵⁴. The ketone body levels are also significantly higher in patients experiencing greater HbA1c improvement bariatric surgery⁵². Ketogenic diets are reported to have favorable effects on energy and glucose homeostasis regulation⁷⁸. Intracerebroventricular injections of 3-hydroxybutyrate in diabetic rats resulted in improved serum glucose levels and peripheral insulin sensitivity due to a potentiated leptin and insulin signaling in the hypothalamus⁷⁸. Thus, earlier increase of 3-hydroxybutyrate days after the bariatric surgery could hypothetically induce a similar central signaling in humans, contributing for a better glycemic control. This particular metabolite could enable the early identification of patients more likely to achieve T2D remission, although with the limitation of not being assessable prior to surgery.

Alanine, pyruvate and lactate decrease after bariatric surgery. This finding was reported to be possibly related to the reduction of gluconeogenic precursors after surgery²². An improved post-surgical mitochondrial function is also suggested, as most of glucogenic AAs that can be converted in Krebs cycle intermediates decrease, while most of Krebs cycle intermediates seems to increase after bariatric surgery. Fasting states or caloric restriction circumstances also generate substrates that upregulate reactions of TCA cycle⁷⁹. Therefore, individuals that experience weight gain are characterized by lower TCA cycle metabolites.

Obesity is also characterized by a blunted post-prandial BAs response. Bariatric surgery, in particular RYGB and BPD normalizes that response, leading to an increase in postprandial BAs that can be justified by the accelerated delivery to the distal intestine where BAs are absorbed^{38, 40, 48}. Since BPD lead to early delivery of food to the more distal gut, BPD would be expected to have a higher effect on BAs levels when compared to RYGB. However, BAs levels were found to be similar after both bariatric surgeries³⁸. Accordingly, bile diversions to the mid-jejunum or mid-ileum led in rats led to the same BAs plasma level, suggesting that BAs absorption is not necessarily proportional to the length of the gut segment⁸⁰. BAs are reported to have a key role on mediating the anti-diabetic effects of the bariatric surgery. Indeed, higher baseline levels of CDCA were correlated with better outcomes after RYGB^{11, 48}. In addition, higher post-operative values of conjugated secondary BAs were associated with an increase of insulin sensitivity after both RYGB and BPD³⁸. BAs metabolic effects are mainly mediated by the activation of two receptors: FXR and TGR5. TGR5 activation in the enteroendocrine cells increase the release of the incretin hormone, glucagon-like peptide-1, known for its anti-diabetic effects⁸¹. In addition, the receptor TGR5 is also expressed in the human muscle and brown adipose tissue, leading to an increase of muscle energy expenditure, via T4-T3 conversion than can further contribute to increase insulin sensitivity^{82, 83}.

Circulating levels of fatty acids, acylcarnitines, phospholipids, triglycerides, total cholesterol, and LDL are frequently elevated in individuals with obesity and particularly with obesity-related metabolic disorders⁵⁷. Bariatric surgery induces an overall improvement in lipid profile with a marked decrease in most lipid classes. Apart from this trend stands the post-operative increase of MCSFA, known for suppressing fat deposition due to enhanced thermogenesis and fat oxidation in animals and humans⁸⁴. Additionally, previous studies also proposed the pharmacological potential of MCSFA for preserving insulin sensitivity in T2D^{85, 86}.

A preoperative lipid profile with higher levels of PES, triglycerides and PLs with long-chain fatty acid could also be useful to assess the likelihood of T2D remission after surgery. Long-chain fatty

acid metabolism is known to be abnormal in diabetes, leading to impaired formation of labeled fatty acids dependent on delta-9, delta-6, and delta-5 desaturation, as well as on chain elongation⁸⁷. Thereby, greater levels of triglycerides with higher carbon number are associated with a decreased diabetes risk⁸⁸. Similar patterns were reported for other lipid classes, including PEs⁸⁹. Palmitic Acid (FFA 16:0) is also strongly associated with T2D, since this main even-chain fatty acid is significantly decreased before¹¹ and after⁴⁹ RYGB in patients with better T2D outcomes. Palmitic acid was shown to activate TLR4 and, subsequently prompt inflammatory cytokines and lipotoxicity in pancreatic β cells, with consequent impaired insulin action⁹⁰.

Although the surgical rearrangements induced by RYGB are more disruptive of normal gastrointestinal anatomy as compared to SG, the alterations induced by the two procedures on the overall lipidomic profile are not dissimilar²⁹. Different findings were observed when the lipidomic profile of the malabsorptive surgery BPD was compared with the one induced by SG, as lipid signatures clearly differ depending on the type of bariatric surgery procedure³⁹. Most sphingolipids and PLs decrease after BPD, which can be attributed to decreased lipid absorption after surgery. On the other hand, PLs levels increased after SG. Circulating levels of PLs are reduced in patients with obesity, which is associated with an increased oxidative status^{91, 92}. SG seems to efficiently restore PLs levels to those found in subjects with normal weight by decreasing patients' oxidative profile. Sphingolipids levels also increase after SG. However, this observation is more difficult to understand since sphingolipids are associated to obesity-related disorders such as insulin resistance and cardiovascular diseases⁹³. Notwithstanding, these lipids have been suggested to be partially associated with the weight regain observed after SG³⁹.

Nonetheless, the interpretation of the summarized data must be evaluated in the context of the limitations of the reported studies. There are some discrepant results across different analysis, raising the possibility of some bias, independent from the anatomical rearrangements of the surgery itself, such as patients' dietary habits, which could also have a significant impact on the metabolome. Additionally, it is important to mention that the time points of assessment before and after surgical interventions differed substantially across the studies, in some cases, the follow-up was not long enough for an adequate appreciation of patients' outcomes. Most of the studies reported the metabolomic profile induced by a single bariatric surgery procedure. Although comparison between studies can be made for understanding the metabolomic profile induced by different surgeries, it raises several constraints derived from the comparison of non-paired patients regarding pre- and post-operative characteristics and from the use of different methodological approaches.

Conclusions | Conclusões

This review summarizes the impact of bariatric surgery on the individuals' metabolomics profile and how different bariatric procedures result in distinct metabolomics signatures, which could contribute to explain the heterogeneity of surgical outcomes. A few pre-operative metabolomics fingerprints were identified as harboring the potential to be used as prognostic biomarkers for weight loss response and T2D remission. Moreover, these studies also highlighted many postoperative metabolic changes that contribute for understanding the molecular mechanisms inherent to dissimilar outcomes. By allowing the prediction of individuals' biological responses, metabolomics studies harbor the potential for identifying those individuals that are more likely to benefit from a given bariatric surgery intervention. Ultimately, this knowledge could positively influence patient care by avoiding unnecessary surgical risks in those that will not benefit from it, towards a more personalized medicine. Ultimately these advances could positively impact the optimization of healthcare resources allocation.

Appendix | Apêndice

 Table I
 Summary of Newcastle-Ottawa Scale scores of the studies included for systematic review

 (cohort studies)
 (cohort studies)

NEWCASTLE-OT	TAWA Q	UALITY /	ASSESSN	MENT S	CALE COHORT S	TUD	IES		
FIRST AUTHOR AND YEAR		SELEC	ΓΙΟΝ		COMPARABILITY	00	тсо	ME	TOTAL SCORE
	1	2	3	4	1	1	2	3	(MAX 9)
Abidi W, 2020 54	0	1	1	1	2	1	1	1	8
Ahlin S, 2019 38	0	1	1	1	1	1	1	1	7
Ahmad NN, 2013 40	1	1	1	1	2	1	1	1	9
Arora T, 2015 ¹⁹	0	1	1	1	2	1	1	1	8
Cabré N, 2020 27	1	1	1	1	2	1	1	1	9
Dadson P, 2020 20	1	1	1	1	1	1	1	1	8
Fiamoncini J, 2018 ²⁶	0	1	1	1	2	1	1	1	8
Friedrich N, 2012 ⁴³	1	1	1	1	2	1	1	0	8
Gralka E, 2015 ²¹	1	1	1	1	2	1	1	1	9
Ha J, 2020 ⁵¹	0	1	1	1	2	1	1	1	8
Herzog K, 2020 35	1	1	1	1	1	1	0	1	7
Hubal MJ, 2017 ²⁸	0	1	1	1	2	1	1	1	8
Jüllig M, 2014 ³¹	0	1	1	1	2	1	1	1	8
Kayser BD, 2017 ⁴²	0	1	1	1	2	1	1	1	8
Khoo CM, 2014 ²²	0	1	1	1	2	1	1	1	8
Kindel TL, 2018 ⁴¹	0	1	1	1	1	1	1	1	7
Kwon HN, 2014 52	0	1	1	1	2	1	0	1	7
Kwon Y, 2020 53	0	1	1	1	2	1	1	1	8
Kwon Y, 2021 ²³	0	1	1	1	1	1	0	1	6
Laferrère B, 2011 ²⁴	0	1	1	1	2	1	1	1	8
Li QR, 2018 50	0	1	1	1	2	1	1	1	8
Lips MA, 2014 ²⁵	1	1	1	1	2	1	0	1	8
Lopes T, 2015 ²⁶	0	1	1	1	2	1	1	1	8
Lopes T, 2016 47	0	1	1	1	1	1	1	1	7
Luo P, 2016 ¹¹	0	1	1	1	1	1	1	1	7
Magkos F, 2013 ¹²	0	1	1	1	1	1	1	1	7
Mendonça Machado M, 2020 ³⁰	0	1	1	1	2	1	1	1	8
Mika A, 2017 ³⁷	0	1	1	1	2	1	1	1	8
Mutch DM, 2009 ¹³	0	1	1	1	2	1	1	1	8
Narath SH, 2016 ¹⁴	0	1	1	1	2	1	1	1	8
Oberbach A, 2011 ³⁴	1	1	1	1	2	1	1	1	9
Ocaña-Wilhelmi L, 2019 32	0	1	1	1	2	1	0	1	7
Palau-Rodriguez M, 2018 ¹⁵	0	1	1	1	2	1	1	1	8

Table I Summary of Newcastle-Ottawa Scale scores of the studies included for systematic review (cohort studies) (cont.)

Ramos-Molina B, 2018 ³⁹	0	1	1	1	1	1	1	1	7
	0	T	1	1		T	T	T	/
Samczuk P, 2018 ²⁹	0	1	1	1	2	1	1	1	8
Samczuk P, 2018 ⁴⁵	0	1	1	1	2	1	1	1	8
Sarosiek K, 2016 ³³	0	1	1	1	1	1	1	1	7
Shantavasinkul PC, 2018 44	0	0	1	1	1	1	1	1	6
Tan HC, 2016 ⁴⁶	1	0	1	1	2	1	1	1	8
Wijayatunga NN, 2018 16	0	1	1	1	2	1	1	0	7
Yao J, 2019 ¹⁷	1	1	1	1	2	1	0	1	8
Yoshida N, 2020 ¹⁸	0	1	1	1	2	1	1	1	8
Yu H 2015 ⁴⁸	0	1	1	1	2	1	1	1	8
Zhao L, 2017 49	0	1	1	1	2	1	1	1	8

Table II Summary of Newcastle-Ottawa Scale scores of the studies included for systematic review(cross-sectional studies)

NEWCASTLE-O)TTAWA S	CALE AI	DAPTED	FOR CR	OSS-SECTIONAL	STUE	DIES	
FIRST AUTHOR AND YEAR		SELEC	ΓΙΟΝ		COMPARABILITY	OUT	COME	TOTAL SCORE
		2	3					(MAX 10)
Jarak I, 2020 56	0	1	1	2	2	2	1	9
Pereira SS, 2020 55	0	1	1	2	2	2	1	9

MAIN FINDINGS	After a mean of 185.3 (72.9) days: increase in total BA con- centrations after both RYGB and BPD.	After 40 weeks: acceleration of the postprandial BA re- sponse, leading to an earlier rise and fall in glycine- and taurine-conjugated BAs after meal ingestion. RVGB normalizes the blunted postprandial circulating BA	After 4 days: reduction in most lipid species. After 42 days: higher levels of decanoic and octanoic acids; increase in most lipid species; lower levels SPMs (18:1/21:0 and 18:1/23:3)	After 12 months: decrease of lefflicine and isoleucine, α- ketoglutarate, 3- hydroxybutyrate, malate, malonil-CoA, glutamate and pyruvate. Increase of Sucinyl-CoA and ox- aloacetate.	After 6 months: decrease in ApoB/ApoA1, BCAA, AAA and GlycA; increase in glycine; lipid parameters remained unchanged.	After 90 days: transient increase in total ACs Sustained de- crease in PCs and increase of SPMs and BAs.	After 3 to 9 days: increase in 3-HB, 2-HB, hippuric acid, trigonelline, and glycine.	After 12 months: increase in arginine, glutamine and dime- thyl sulfate; decrease in AAAs and BCAAs; normalization of pyruvate, methanol and isopropanol; temporary increase of citrate. TMAO increased 12 months after SG only.
METABOLOMIC ANALYSIS	Targeted UPLC- MS	Targeted UPLC- MS	UPLC-MS	Targeted GC-MS and UHPLC	H NMR	Targeted LC- MS/MS (lipidomic)	H NMR	Untargeted H-NMR
BIOLOGICAL SAMPLE	Fasting plasma samples	Fasting and postprandial plasma samples	Fasting plasma samples	Fasting plasma samples	Fasting serum samples	Fasting plasma and dried blood spots samples	Urine	Fasting serum
POST-OPERATIVE BMI (KG/M²)	37.7 ± 7.4	31.5±5.0	Reduction of 31.3 ± 2.1%	NA	31.8 ± 13.5	35 ± 3.3	NA	31.7 ± 4.1(SG) 29.4 ± 5.9 (proximal RYGB) 30.7 ± 4.9 (distal RYGB)
FEMALE: MALE	1:2	1:4	11:5	67:23	23:0	18:8	NA	77:33
AGE AT SUR- GERY (YEARS)	44.3 ± 8.3 (SD)	44.8±12.9	47.4±1.9	Median: 49 (41-58)	42.8±9.6	36.2 ± 7.8	Range: 22 to 63	43.6±1.0
PRE-OPERATIVE BMI (KG/M²)	51.6±9.6 (SD)	47.7 ± 7.4	48.9±1.3	Median: 46.4 (42.4-51.6)	41.1±4.2	44.1±3.6	Median: 48.3 (42.9-52.5)	46.2 ± 7.8
SUBJECTS UNDER SURGERY	N=15	N=5	N=16	N=270	N=23	N=26	N=50	N= 106
SURGICAL PROCEDURE	RYGB and BPD	RYGB	RYGB	NA	RYGB and SG	RYGB	RYGB and SG	SG, proximal RYGB and distal RYGB
EXPERIMENTAL DESIGN	Prospective co- hort study	Prospective co- hort study	Prospective co- hort study	Prospective co- hort study	Prospective co- hort study	Prospective co- hort study	Prospective co- hort study	Prospective co- hort study
FIRST AUTHOR AND YEAR	Ahlin S, 2019 ³⁸	Ahmad NN, 2013 ⁴⁰	Arora T, 2015 ¹⁹	Cabré N, 2020 ²⁷	Dadson P, 2020 ²⁰	Fiamoncini J, 2018³ ^{sí}	Friedrich N, 2012 ⁴³	Gralka E, 2015 ²¹
	EXPERIMENTAL SURGICAL UNDER UNDER BMI UNDER BMI DESIGN PRE-OPERATIVE AGE AT SUR. FEMALE: POST-OPERATIVE BIOLOGICAL SAMPLE MATABOLOMIC ANALYSIS DESIGN PROCEDURE (KG/M)	EXPERIMENTAL SURGICAL SUBJECTS PRE-OFERATIVE BMI GER AT SUR- BMI FEMALE: POST-OFERATIVE BMI (KG/M*) METABOLOMIC METABOLOMIC DESIGN PROCEDURE SURGERY (KG/M*) GERY (YEARS) MALE BIOLOGICAL SAMPLE ANALYSIS Prospective co- hort study RYGB and BPD N=15 51.6.4.9.6 44.3.4.8.3 1.2 37.7.4.7.4 Fasting plasma samples Targeted UPLC- MS	EXPERIMENTAL DESIGN SURGICAL surgers PRE-DERATIVE BMI GEN (FEAAS) GEN (FEAAS) FMALE: BMI (KG/M) DOST-OFERATIVE BMI (KG/M) RIPABOLOWIC METABOLOMIC DESIGN UNDER SURGERV UNDER SURGERV UNDER BMI BMI (KG/M) BMI (KG/M) BIOLOGICAL SAMPLE AMAYSIS Prospective co- hort study MYGB and BPD N=15 51.6.4.9.6 44.3.4.8.3 1.2 37.7.4.7.4 Fasting plasma samples Targeted UPLC- MS Prospective co- hort study RYGB N=5 47.7.4.7.4 44.8.1.2.9 1.3 31.5.4.5.0 Fasting plasma samples Targeted UPLC- MS	EXPERIMENTAL DESIGNSURGICAL SURGERYPRE-DERATIVE BMI CERV (FGARS)GER (FEAALE BMI (FG(AF))DICOGICAL SAMPLEMETABOLOMIC ALAYSISProspective co- hort studyWGB and BPDN=15S16 ± 9.644.3 ± 8.31.2377 ± 7.4BIOLOGICAL SAMPLEARAYSISProspective co- hort studyWGB BPDN=15S16 ± 9.644.3 ± 8.31.2377 ± 7.4Fasting plasma samplesTargeted UPLC- MSProspective co- hort studyWGBN=547.7 ± 7.444.8 ± 12.91.431.5 ± 5.0Fasting and postprandialTargeted UPLC- MSProspective co- hort studyN'GBN=547.7 ± 7.444.8 ± 12.91.431.5 ± 5.0Fasting and postprandialTargeted UPLC- MSProspective co- hort studyN'GBN=1643.8 ± 12.91.431.5 ± 5.0Fasting and postprandialTargeted UPLC- MSProspective co- hort studyN'GBN=1643.8 ± 12.91.431.5 ± 5.0Fasting and postprandialTargeted UPLC- MSProspective co- hort studyN'GBN=1643.8 ± 12.91.431.5 ± 5.0%Fasting plasma samplesGCMS and MS	DYSERMENTIAL DESIGNUNECT BOL COUNCRRELOPERATIVE BMI COUNCRRELOFERATIVE BMI (GGN)GER VEANS BMI (GGN)MALE BMI (GGN)DOCOLOL SAMPLE BMI (GGN)MALTSS ALLSSProspective co- bor studyRYGB and BPDN=1551.6.19.644.3.3.8.31.237.7.17.4Fasting plasma samplesTargeted UPLC- MSProspective co- bor studyRYGBN=1551.6.19.644.3.1.8.31.231.7.17.4Fasting plasma samplesTargeted UPLC- MSProspective co- bor studyRYGBN=547.7.17.444.8.1.12.91.431.5.5.0Fasting plasma samplesTargeted UPLC- MSProspective co- bor studyRYGBN=547.4.1.91.41.4.8.1.12.91.481.5.5.0Fasting plasma samplesTargeted UPLC- MSProspective co- bor studyRYGBN=547.4.1.91.1.581.5.1.5.0Fasting plasma samplesGGMS and UPC.MSProspective co- bor studyNN=270Median: 46.447.4.1.91.1.581.3.2.1.96Fasting plasma samplesGGMS and UPC.MSProspective co- bor studyNNNFasting plasma samplesTargeted UPLC- MSProspective co- bor studyNNFasting plasma samplesTargeted UPLC- MSProspective co- bor studyNNFasting plasma samplesTargeted UPLC- MSProspective co- bor studyNNNFasting plasma samplesTargeted UPLC- MSProspective co- <th>Definition Deficiency Description Description Description Description DescriptionDescription Description Description Description DescriptionDescription Description Description Description DescriptionDescription Description Description DescriptionDescription Description Description DescriptionDescription Description DescriptionDescription Description DescriptionDescription Description DescriptionDescription </br></th> <th>DEGRAMENTAL DEGRAMENTAL BURCEDSUBJECTS LABED LUNCEDFRE-OFREATING LABEDFRE-OFREATING LANT LABEDFRE-OFREATING LANT LANTFRE-OFREATING LANT LANT LANT LANT LANTEUGC COLAMANE LANT L</th> <th>Desimetively Desider<</th>	Definition Deficiency 	DEGRAMENTAL DEGRAMENTAL BURCEDSUBJECTS LABED LUNCEDFRE-OFREATING LABEDFRE-OFREATING LANT LABEDFRE-OFREATING LANT LANTFRE-OFREATING LANT LANT LANT LANT LANTEUGC COLAMANE LANT L	Desimetively Desider<

	25)						טוובט טו אמיירויי		
Prospective co- hort study	ģ	RYGB	N=19	39.8±3.3	43 ± 6.3	0:61	33.3 ± 3.1	Fasting and postprandial plasma	UPLC	After 1 day: reduction of medium-chain ACs and purines; increasing in short-chain AC 2:0, most aminoacids, car- nitine and 3-HB; few lipids were altered. After 6 weeks: reduction in 6 out of the 20 detected ACs and 3-HB; few lipids were altered.
ospective c hort study	Prospective co- hort study	GB	9=V	51.2±8.8	38.5 ± 6.8	6:0	32.6±8.1	Plasma and serum sam- ples	LC-MS	After 1 year: increased levels of apargine, cirtulline, gluta- mine, glycineand histidine; lower levels of cysteinylglycine, glutamic acid, leucine/isoleucine, total BCAAs and Glu/Gln ratio.
s <u>çi</u>	Prospective co- hort study	RYGB and SG	N=15	42.1±4.0 (RYGB) 42.3±5.9 (SG)	41.0 ± 3.1 (RYGB) 46.8 ± 2.9 (SG)	14:1	NA	Fasting plasma samples	Untargeted GC-MS	3 days after RYGB: decrease in histidine, proline, citrate and decanoic acid. 3 days after SG: increase in 2-HB and 3-methyl-2-oxo-pen- tanoic acid.
s gi	Prospective co- hort study	RYGB and LAGB	N=59	46.5 ± 1.0 (RYGB) 43.6 ± 0.7 (LAGB)	37.3 ± 1.9 (RYGB) 34.5 ± 1.6 (LAGB)	59:0	38.0 ± 1.2 (RYGB) 38.3 ± 1.0 (LAGB)	Fasting serum samples	Targeted LC-MS/MS (lipidomic)	After 3 months, the majority of lipids decreased after both surgical procedures.
ਣ ਦ	Prospective co- hort study	RYGB	N=20	45.6±2.4	47.9±5.0	2:2	Reduction of 6.5 ±1.0%	Fasting and postprandial plasma	Targeted tandem MS	10 to 14 days after, in fasting analysis: - reduction in AA (profine, histidine, valine, phenylalanine, BCAA, AAA and total AA), alanine, and molar sum of C3 and C5 ACS; increasing in C2, long-chain (C14-C22), total AC, NEFA, ketones and 3-HB. 10 to 14 days after, in postprandial analysis: - reduction in multiple AA, molar sum of BCAA, aromatic and total AA; increasing in plasma C2, medium-chain, long- chain and total ACs
2 S	Prospective co- hort study	SG	N=28	45.0±6.8	45.4 ± 12.9	41:9	38.9±6.3	Fasting and postprandial serum samples	NPLC-MS	After 12 weeks, increase in multiple BA subtypes: fasting HCA (total, unamidated and G- sub-fractions), postprandial BAS (total and G-), CDCA (total and G), postprandial MCA (total and G-) and postprandial HCA (to- tal, unamidated and G-)
D N	Prospective co- hort study	RYGB	N=23	38.9±5.2	41.8±13.1	17:6	31.4±5.5	Fasting serum samples	Targeted LC–MS	After 3 months: decrease in KynP metabolites, BCAAs, AAAs and TDGMs.
D v G	Prospective co- hort study	GB	N=21	44.9±8.7 (SD)	43.3 ± 10.0 (SD)	NA	Decrease of 4.60 ± 2.13	Fasting plasma samples	Targeted tandem MS	After a BMI decrease of 4.60 \pm 2.13 kg/m² in BMI, individuals had a significant decrease in AAs, in particular BCAAs and related metabolites.

				43.1 ± 0.9 (LAGB)	46.3 ± 1.9 (LAGB)		39.0 ± 0.8 (LAGB)			
Lips MA, 2014 ²⁵	Prospective co- hort study	RYGB and LAGB	N=27	44.2 ± 0.8 (RYGB and no T2D)	48.6 ± 1.6 (RYGB and no T2D)	27:0	36.63 ± 0.8 (RYGB and no T2D)	Fasting plasma samples	Targeted UPLC - tandem MS (AA analysis)	After 3 months: significant decrease in BCAAs in both pro- cedures
				43.5± 1. 1 (RYGB and T2D)	51.3 ± 1. 9 (RYGB and T2D)		34.7 ± 0.8 (RYGB and T2D)			
Lopes П, 2015 ²⁶	Prospective co- hort study	RYGB	N=10	32.38 ± 2.11	Range: 25 to 65	1:1	25.48 ± 1.85	Fasting and postprandial plasma samples	H NMR and CG-MS	After 12 months: decrease in lactate, BCAAs, very low-den- sity lipoprotein, low-density lipoprotein, N-acetyl- glyco- proteins, and unsaturated lipid; increase in PCs and HDL
Luo P, 2016 ¹¹	Retrospective cohort study	RYGB	N=35 ^s	30.8±3.3	49.8±9.9	19:16	24.3±2.3	Fasting serum samples	Untargeted UPLC-MS	After 6 months:77 altered metabolites. After 12 months: 88 altered metabolites (64 common with 6 months). Alterations (mainly decrease) were observed in: AA and their derivates, FFAs, ACS, BAs, LPCs, PCs and SPMs.
Magkos F, 2013 ¹²	Prospective co- hort study	RYGB and LAGB	N=20	45.6 ± 6.7 (RYGB) 46.5 ± 8.8 (LAGB)	43 ± 7 (RYGB) 47 ± 14 (LAGB)	17:3	36.4±5.0 (RYGB) 37.6±7.3 (LAGB)	Plasma samples	MS/MS (AAs and ACs analysis)	Post-operative decrease levels of BCCAs, C3 and C5 acyl-carnitine in both surgeries (22 \pm 7 weeks after RYGB and 16 \pm 2 weeks after LAGB)
Mendonça Machado N, 2020 ³⁰	Prospective co- hort study	RYGB	N=28	Weight: 112.8 ± 15.6 Kg	Range: 18–60	28:0	Weight: 91.8±12.2 Kg	Plasma samples	Untargeted GC-MS	After 3 months: decrease levels of dicarboxylic acids (aminomalonate, fumaric acid, malic acid, oxalic acid); en- riched metabolic pathways included arginine and proline metabolism, urea and TCA cycles, gluconeogenesis, malate-aspartate shuttle, and carnitine synthesis.
Mika A, 2017 ³⁷	Prospective co- hort study	RYGB, OLGB and SG	N=16	41±1.1	44 ± 3.1	NA	31±1.1	Serum samples	H-NMR	After 6 months: significant decrease in the levels of all analyzed lipids - TGs, PLs (PCs, Pes and SPMs), total, free and esterified cholesterol, total and specific fatty acids. The most evident decrease was in 7-lathosterol.
Mutch DM, 2009 ¹³	Prospective co- hort study	RYGB	N=14	46.2±1.7	45.4±3.6	14:0	35.1±1.7	Serum samples	Untargeted GC- MS and LC-MS	After 6 months: decrease levels of BCAAs, ceramide, TGs and saturated fatty acids; increase levels of specific SPMs, unsaturated fatty acids and PLs.

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	RYGB	N=44	43.9±5.4 (SD)	46.8±11.3 (SD)	29:15	30.0±4.4 (SD)	Serum samples	Untargeted LC- HRMS	After 1 year: increase levels of TMAO indoxyl-sulfate, gly- cine and PC C40:7; decrease levels of BCAAs, choline, tyro- sine, alanine and phenylalanine. V pattern (only decrease at short time) observed for Crea- tine, LysoPC C16:1 and c18:2, Ornithine, PC C34:3, PC C36:5, PC C36:6, Sarcosine, Tryptophan and Uracil. A pattern (only increase at short time) observed for Acetyl- glycine, Arginine, Camitine, Hydroxyisobutyric acid, Leu- cine Proline, Pantothenic acid, PC C38:6, Pyroglutamic acid and Threonine.
	SG	N=14	54.00 ± 8.05	41.9±13.29 (SD)	9:5	36.31 ± 7.25 (SD)	Fasting serum samples	Targeted MS/MS	After 6 months: increase in glutamine; decrease in phos- phatidylcholine diacyl C42:0 and C18:2 carnitine.
	SG	N=32	50.4 ± 7.2 (SD)	44.6 ±7.6 (SD)	25:7	38.1±5.9 (SD)	Fasting serum samples	Targeted UPLC- MS/MS	After 6 montths: increase levels of polyamide metabo- lome: putrescine, acetyl-spermidine (N8-acetyl-spermi- dine, and N1,N8-diacetyl-spermidine), and N1,N12-diace- tyl-spermine.
	SG and RYGB	N=39	50.52 ± 8.37	Range: 19 to 59	27:12	36.42 ± 6.14	Fasting serum samples	Untargeted LC-MS	After 1, 3 and 6 months: alteration in the indoles and deri- vates, AAs, glycerolipids, glycerophospholipids and fatty acids were observed.
	SG and BPD	N=37	47.9±6.1 (SG) 41.8±6.9 (BPD)	47.0 ± 6.7 (SG) 44.4 ± 8.2 (BPD)	22:15	36.5 ± 4.5 (SG) 39.7 ± 4.4 (BPD)	Fasting plasma samples	UPLC-MS (lipidomics)	After 6 months: BPD induced an overall decline in sphin- golipids and PLss and an increase in BAs levels; SG induced an increase of sphingolipids and PLs, and no changes in BAs levels
Prospective co- hort study	SG and RYGB	N=54	50.92 ± 7.33 (SG) 45.79 ± 5.5 (RYGB)	49.3 ± 8.7 (SG) 50.1 ± 9.3 (RYGB)	29:25	37.33 ± 7.3 (SG) 32.61 ± 5.5 (RYGB)	Fasting serum samples	Untargeted GC- MS and LC-MS	6 months after both procedures: decrease in PCs, IPCs, Pes and LPEs; increase in SPMs and choline. After RYGB only: increased levels of sulfate containing me- tabolites. After SG only, increased p-cresol.

				48.74 ± 8.2 (SG with T2D)	46.0 ± 12.84 (SG and T2D)				Intargeted	After 28 days: increased levels of histidine, trans-
Sarosiek K, 2016 ³³	Prospective co- hort study	SG and RYGB	N=15	43.54 ± 4.13 (SG and no T2D)	45.2 ± 12.24 (SG and noT2D)	12:3	NA	Fasting serum samples	UPLC-MS/ MS and GC-MS	u ocaliate, us unocaliate, pyrogutallyvallie, liente, guta- thione, and its precursors, pentose phosphate pathway in- termediates, derivate pentose sugars and 3-phospho- elverate. Derease in ascarbate, various toconherols and
				47.56 ± 6.61 (RYGB)	44.4±17.57 (RYGB)					pyruvate.
Wijayatunga NN, 2018 ¹⁶	Prospective co- hort study	RYGB	N=20	46.83±6.21	37.25 ± 11.68	15:5	34.34 ±6.44	Fasting serum samples	Untargeted GC- MS and NMR	After 6 months: decrease on BCAAs, 2-aminobutyrate, bu- tyrate, 2-HB, 3-HB. acetone, 2-methylglutarate, and 2-ox- oisocaproate; increase in alanine, glycine, pyruvate, tau- rine and fatty acids (C10:0, C13:0, C14:0, C15:0, and C18:0).
Yao J, 2019 ¹⁷	Prospective co- hort study	SG	N=11	39.1±1.4	36.8±9.1	8:3	31.5 ± 1.1	Plasma samples during insulin clamp and fasting	NA	After 6 months: decrease in BCAAs, phenylalanine, glycine and tyrosine levels (during fasting and insulin damp); de- crease in methionine and glutamate/glutamine concentrations (dur- ing insulin damp).
Yoshida N, 2021 ¹⁸	Prospective co- hort study	SG	N=15	40.8±6.6	52.2±6.5	12:3	33.5±6.1	Fasting plasma samples	Untargeted MS	After 3 months, decrease in BCAAs, tryptophan, phenylala- nine, tyrosine, choline, creatine, orithine, hypoxanthine, 2-oxoisovaleric acid, 2-HB, N,N-dimethylglycine and uri- dine. Increase in citric acid, succinic acid, malic acid, argi- nine and glycine.

- Chenodeoxycholic acid; DD8 - Duodenal-jejunal bypass; FFA - Free fatty acids; G-- glycine amidated; GB - Gastric bypass; GC - Gas chromatography; Glu/Gln - glutamic acid/glutamine; H-NMR - Proton nuclear magnetic resonance; HCA -Hyocholic acid; HRMS - High resolution mass spectrometry; IR - Insuline Resistance; KynP - Kynurenine pathway; L4GB - Laparoscopic adjustable gastric band; LC - Liquid chromatography; LPC - Lysophosphatidylcholines; LPE - Lysophosphatidylethanolamines; MCA -Beta-muricholic acid; MS - Miss Spectrometry; IR-Insuline Resistance; OLGB - Omega-loop gastric bypass; PC- Phosphatidylcholines; PE - Phosphatidylcholines; PL - Phosphatidylcholines; PL - Phosphatidylcholines; PL - Phosphatidylcholines; PL - Phosphatidylcholines; PC - Steve gastrectomy; SPM -Sphingomyelin; T2D – Type 2 diabetes; TCA - tricarboxylic acid; TDGM - tryptophan-derived gut microbial metabolites; TG - Triglycerides; UPLC - Ultra-Performance Liquid Chromatography: 5 - Salan cohort. Abbreviations: 2-HB - 2-hydroxybutyrate; 3-HB - 3-Hydroxybutyrate; AAA - Aromatic amino acid; AA - Amino acid

FIRST AUTHOR AND YEAR	EXPERIMENTAL DESIGN	SURGICAL PROCEDURE	SUBJECTS	PRE- OPERATIVE BMI (KG/M ²)	AGE AT SURGERY (YEARS)	FEMALE: MALE	POST-OPERATIVE BMI (KG/M²)	BIOLOGICAL SAMPLE	METABOLOMIC ANALYSIS	OUTCOMES AND MAIN FINDINGS
			Control group -	98+227	42.0 ± 11.1		Nadir BMI: 28.2 ± 4.8			Outcomes: Patients with sustained weight loss at 5.0 \pm 3.7 years (N=14) and patients with weight regain at 8.9 \pm 3.0 (N=21)
Abidi W,	Prospective		patients with obesity (N=11)	0.0 I ///4			Current BMI: 29.4 ± 4.6	Serum fasting		Patients with sustained weigh loss: Depletion in metabolites of fatty acids metabolism, including TGs, DGs
2020 ⁵⁴	cohort study	КУСВ	Patients with				Nadir BMI: 28.9 ± 2.9	samples	LC-MS	and CLs; increased levels or beta-hydroxyburyrate and gycine Patients with weigh regain
			obesity undergoing RYGB (N=35)	49.9±7.3	47.9 ± 10.7	1:07	Current BMI: 37.9 ± 5.7			Decreased levels of metabolites related to serine, giyone and threonine pathway, phenylalanine metabolism, TCA cycle, and alanine and glutamate metabolism, higher levels of other AAs.
Ahlin S,	Prospective	RYGB + BPD	Patients with obesity (N= 15), in	51.6±9.6	44.3 ± 8.3	1:2	37.7 ± 7.4	Fasting	Targeted	Outcome: Patients with obesity and improved IR after a mean of 185.3 (72.9) days
~F107	conort stuay		wnicn 13,3% with T2D (N= 2)	(US)	(US)			plasma	UPLC-MS	Increase in total BAs concentrations after RYGB and BPD.
							Bodinetion of			Outcomes: patients with (N=7) or without (N=7) T2D remission at 2 years
			Dationts with	51.0±2.3	48.1±2.3	3:4	30.5 ± 4.4 %			Patients with T2D remission:
Arora T, 2015 ¹⁹	Prospective cohort study	RYGB	obesity (N= 16), in which 87.5% with					Fasting plasma	GC-MS and UPLC-MS	Before surgery, higher levels of metabolites from TCA cycle and pentose phosphate pathways, TG and PLs with LCFAs, and PEs.
			T2D (N= 14)	47.2±1.6	48.8 <u>+</u> 3.5	7:0	Reduction =			After 4 days, higher levels of aconitate, indole acetic acid and ribitol; reduction in most lipid species.
							2			After 42 days, higher levels of decanoic and octanoic acids; increase in most lipid species.
				39.6 ± 7.9			28.8±6.6			Outcomes: patients with (N=14) or without (N=10) T2D remission at 1 year
			Dationto mith	(SD)						Patients with T2D remission:
Ha J, 2020 ⁵¹	Prospective cohort study	RYGB + SG	obesity ⁵ and T2D (N= 24)		45.4 ± 10.2 (SD)	17:7		Fasting serum samples	Targeted LC- MS	Higher baseline levels of L-DOPA and 3-HAA (sustained up to 3 months after surgery).
				33.9 ± 4.5 (SD)			28.9 ± 5.6			At 1 year, no differences in the levels of L-DOPA and 3-HAA between both outcomes.
										No distinction between both surgery procedures.

Table IV Summary of studies comparing the metabolomic profiles of patients with different outcomes (T2D and weight loss)

	Outcomes: Patients with (N=23) or without (N=I2) T2D remission at 1 year Significant metabolic alterations related to FFA, acylcarnitines, AAs, BAs, and lipids species in both outcomes.	Patients with T2D remission: Higher baseline levels of glycoursodeoxycholate sulfate, bilirubin and tryptophan.	6 months after RYGB: significant decrease in FFA 16:0, FFA 18:3, FFA 17:2 and hippuric acid.	Outcomes: Patients with high or low WL at 1 year (above or below the median of the WL)	After 1 year, significant decreased levels of creatine, ornithine, arginine and valine in patients with high WL.	Outcomes: Patients with quicker (N=11) or slower (N=9) remission of T2D	Quicker responders:	Lower baseline levels of most LPC and LPE.	Postoperative significantly changed metabolites can be classified mainly as PC, LPC, PE, LPE, AAs, organic acids, sugars and metabolites related to gutmicrobiota metabolism.	Outcomes: Patients with weight loss and improved HOMA-IR after RYGB (N=11) and patients with similar weight loss after WLMP (N=11)	Patterin Long Rough X. 2009. Higher pre-operative levels of 2-HB. After 6 months, greater decline of 2-HB, 3-HB, acettacetic acid, CMPF, and hyposarthine proventine and hyposarthine After 6 months, greater decline of BAAs and significant correlation between improved HOMA-IR and decline of valine and BCAAs.	Outcome: patient with weight loss and improved IR resistance at 6 months after SG (N=8)	Patients undergoing <u>5G:</u> After 6 months, reduction in total BCAAs, acyteamitines (C3 and C4) and leucine oxidation.
	Sigr		0	Ou Untargeted		0	Untargeted GC-MS and	LC-MS	Pos as F	Out	GC/MS and Af Af	(Targeted GC- Mcc.md1C-	
	Serum	samples		Serum	saiduas		Fasting serum				samples	Fasting	samples
•	24.3±2.4		24.2±2.3	At 1-3 weeks: 40.8 ± 5.2 (SD)	At 1 year: 30 ± 4.4 (SD)		3/±3.b		43 ± 7.6	Decrease of 7.5±1.2	Decrease of 8.9 ± 1.9	W	Median: 30.5 (29.1–32.6)
		19:16		29:15		Ş	4:1	T	4:5	4:7	8:3	0:10	7:3
	49.8± 9.9 (6months	after surgery)		46.8 ± 11.3	(nc)		4/ ± 10.3		51 ± 11.4	56.5 ± 11.4	50.5 ± 7.8	Median: 30.3 (25.1–36.1)	Median: 29.5 (26.8–41.8)
)		30.8 ± 3.3		43.9±5.4 /201	(רוכי)		49 ± 4.5		51±8.3	38.2 ± 4.6	41.3±3.0	Median: 23.2 (22.1–23.7)	Median: 38.5 (37.0–40.4)
	Patients with	obesity ⁵ and T2D (N=35)		Patients with obesity (N= 44), in	T2D (N= 24)		Patients with obesity and T2D	(N=20)		Control group - patients with obesity in a WLMP (N=11)	Patients with obesity and T2D undergoing RYGB (n=11)	Control group - patients with normal weight (N=10)	Patients with obesity and IR undergoing SG (N=8)
		RYGB		RYGB			SG				RYGB	ទ	R
	Retrospective	cohort study		Prospective	corror study		Prospective	cohort study			cohort study	Prospective	cohort study
	Luo P,	2016 ¹¹		Narath SH,	BTN7		Samczuk P,	2018-2		ł	PC, 2018 ⁴⁴	Tan HC,	2016 ⁴⁶

Table IV Summary of studies comparing the metabolomic profiles of patients with different outcomes (T2D and weight loss) (cont.)

iable IV summary of studies comparing the metabolomic profiles of patients with different outcomes (1 2D and weight loss) (cont.)	Outcome: Patients with significant weight loss SG induced an early (6 weeks after) and persistent (12 weeks after) increase in multiple BAs subtypes: -fasting HCA (total, unamidated and G- sub-fractions). post-prandial BAs (total and G-), CDCA (total and G), MCA (total and G-), HCA (total and G-), CDCA was significantly correlated to Increased post-prandial total and G-CDCA was significantly correlated to increased weight loss at both 6 and 12 weeks.		After 1 week, 3-HB is higher in the improved group, whereas glucose and lipids (LDL, VLDL) are higher in the non-improved group. No distinction between both surgery procedures.	Outcome: Patients with significant weight loss Dutcome: Patients with significant weight loss Before SG, Jower levels of S-HIAA, S-HIAA/serotonin ratio and BCAA (++isoleucine), higher levels of S-HIAA, S-HIAA/serotonin ratio and BCAA (++isoleucine), were significantly associated with greater WL% at 3 and 6 months after SG.	Outcome: Patients with obesity and T2D, but improved IR at 3 months KynP metabolites levels were higher before surgery and decrased after G. BCAAs and AAAs levels decreased after surgery. TDGMs decreased after surgery.	Outcome: Patients with obesity and no T2D (N=10) and patients with obesity and T2D, but improved IR at 12 months (N=9) IS After RYGB, significant decrease in fasting plasma BCAAs was associated with improved IR.	Outcome: Patients with T2D remission at 1 year After RYGB: Iower levels of BCAAs. After RYGB in postprandial analysis: decreased VLDL, LDL, N-acetyl glycoproteins and unsaturated lipids; increased HDL and PC.
anterer	NPLC-MS	GC-MS and	H-NMR	Targeted LC- MS	Targeted LC- MS	LC-MS/MS	H-NMR
ופנורצ אורנו	Fasting/postpr andial serum samples	Plasma	samples	Fasting serum samples	Fasting serum	Fasting and postprandial plasma	Plasma samples
lies or par	At 6 weeks: 40.7 ± 6.1 At 12 weeks: 38.9 ± 6.3	27.2 ± 3.0	21.7±2.2	At 3 months: 31.5 ± 5.1 At 6 months: 27.9 ± 4.6	31.4±5.5	31.0±5.1 (no T2D) 33.9±7.5 (T2D)	25.48 ± 1.85
піс ргоі	41:9	1:1	7:3	17:10	17:6	NA	1:1
ופוטמשפו	45.4±12.9	39.8 ± 9.2	49.8 ± 6.9	42.1±12.9	41.8±13.1	43±13 (no T2D) 52±9 (T2D)	25 to 65
ing the fi	45.0±6.8	30.8 ± 5.6	24.6±3.1	38.7 ± 5.2	38.9 ± 5.2	41.5 ± 4.8 (no T2D) 43.1 ± 5.1 (T2D)	32.38 ± 2.11
uales compar	Patients with obesity (N= 28), in which 21% with T2D	Patients with	(N= 22)	Patients with obesity (N= 27), in which 74% with T2D (N=20)	Patients with obesity and TZD (N=23)	Patients with obesity (N=19), in which 47% with T2D (N=9)	Patients with obesity and T2D (N=10)
iary or su	S	an - 200		SG	SG	RYGB	RYGB
	Prospective cohort study	Prospective	cohort study	Prospective cohort study	Prospective cohort study	Prospective cohort study	Prospective cohort study
ומטו	Kindel T, 2018 ⁴¹	Kwon H,	2014 ⁵²	Kwon Y, 2020 ⁵³	Kwon Y, 2021 ²³	Li QR, 2018 ⁵⁰	Lopes T, 2016 ⁴⁷

Table IV Summary of studies comparing the metabolomic profiles of patients with different outcomes (T2D and weight loss) (cont.)

Outcomes: Patients with (N=26) or without (N=12) T2D remission at 2 years <u>Patients with TD2 remission:</u>	Higher baseline levels of CDCA relative to the total BAs. <u>Patients with noTD2 remission:</u> 1 year after RYGB, total BAs were significantly lower.	Outcomes: Patients with (N=26) or without (N=12) T2D remission at 2 years	Patients with T2D remission: Higher baseline ratios of stearic/palmitic acid (C18:0/C16:0) and eicosadienoic/linoleic acid (C20:2 n6/C18:2 n6).
Targeted	UPLC	Targeted UPLC-	di UFMS (free fatty acids)
Fasting serum	samples	Fasting serum	samples
24.9±2.9	24.9±2.5	24.4±2.8	24.8±2.4
11	7:5	1:1	7:5
437+128	0.91 1	43.7 ± 12.8	
32.8±3.9	31.0±3.5	32.8 ± 3.9	31.0 ± 3.5
Patients with	(N=38)	Patients with obesitv ⁵ and T2D	(N=38)
asya		RYGB	
Retrospective	cohort study	Prospective	conort study
,Ни Ү *	2015 ⁴⁸	*Zhao L,	-/107

Table IV Summary of studies comparing the metabolomic profiles of patients with different outcomes (T2D and weight loss) (cont.)

DCA - Deoxycholic acid; DG - Diglyceride; DJB - Duodenal-jejunal bypass; FFA - Free fatty acids; G - Gycine amidated; GC - Gas chromatography; H-NMR - Proton nuclear magnetic resonance; HCA - Hyocholic acid; HRMS - High resolution mass spectrometry; * - Both studies had the same participants; \$ - Asian cohort. Abbreviations: 2-HB - 2-hydroxybutyrate; 3-HAA - 3-hydroxybutyrate; 3-HB - 3-hydroxybutyrate; 5-Hydroxybutyrate; 5 IR - Insuline Resistance; KynP - Kynurenine pathway; L-DOPA - L-dihydroxyphenylalanine; LC - Liquid chromatography, LCFA - Long-chain fatty acid; LDL - Low density lipoprotein; LPC - Lysophosphatidylcholines; LPE - Lysophosphatidylcholines; L amino acid; AA - Amino acid; BA - Bile acid; BCA - Branched chain amino acids; BMI- Body mass index; BPD – Biliopancreatic diversion; CDCA - Chenodeoxycholic acid; CE - Cholesterol ester; CMPF - 3- carboxy-4-methyl-5-propyl-2-furanpropionic acid; - Beta-muricholic acid; MS - Mass Spectrometry; NGT - Normal glucose tolerance; PC - Phosphatidylcholines; PE - Phosphatidylethanolamines; PL - Pho Tricarboxylic acid; TDGM - Tryptophan-derived gut microbial metabolites; TG - Triglyceride; UPLC - Ultra-Performance Liquid Chromatography; VLDL - Very low density lipoprotein, WL - Weight loss; WLMP - Weight loss Maintenance Program

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FIRST AUTHOR AND YEAR	EXPERIMENTAL DESIGN	PARTICIPANTS	SURGERY GROUPS	AGE AT SURGERY (YEARS)	FEMALE: MALE	PRE-OPERATIVE BMI (KG/M ²)	TIME AFTER SURGERY	POST- OPERATIVE BMI (KG/M²)	SAMPLE TYPE	METABOLOMIC METHOD	MAIN FINDINGS
						LAGB vs RYGB	YGB				
Magkos F, 2013 ¹²	Prospective cohort	Patients with obesity and NGT	LAGB (n=10)	47±14	1:6	46.5±8.8	22±7 weeks	37.6±7.3	Fasting	Targeted MS/MS (AAs and	BCAAs and acylcarnitines decreased after both
	study	(N=20)	RYGB (n=10)	43±7	8:2	45.6±6.7	16±2 weeks	36.4 ± 5.0	plasma	acylcarnitine)	surgery in similar proportions
		Patients with	LAGB	017637	0.11	007167	3 weeks	41.10 ± 0.85			BCAAs levels decreased after both surgeries.
Lips M, 2014 ²⁵	Prospective cohort	obesity and NGT (N = 74: n= 27	(n=11)	C'T I C'04	0.111	C.O.I. T.C.	3 months	39.02 ± 0.82	Fasting	Targeted UPLC- tandem MS	BCAAs levels decreased was higher in patients submitted to RYGB.
	study	submitted to bariatric surgeries)	RYGB	217200	0.91	007677	3 weeks	40.48 ± 0.85	piasma	(AAs analysis)	Glycine and Serine levels only increased after RYGB.
			(n=16)	0.T I 0.04	D:0T	44.4 I U.O	3 months	36.63±0.82			AAAs levels only decreased after RYGB.
		Patients with	LAGB	315476	0.66	436407	1 month	40.4 ± 1.0			The majority of lipids decreased equivalently between both surgical
Kayser BD,	Prospective cohort	obesity and at least one severe	(n=22)	0.1 - 0.40	0.777	10-1 0-Ct	3 months	38.3±1.0	Fasting	Targeted LC-	groups.
-/107	study	obesity-related comorbidity	RYGB	373+19	37-0	465+10	1 month	41.1±1.1	serum	MS/MS (lipidomic)	Some PC and SM species decreased 1 month after surgery and remained suppressed 3 months after RYGB, while they either returned
		(YC=N)	(n=37)		2		3 months	38.0±1.2			or tended to return to baseline values 3 months after LAGB.
						SG vs RYGB	GB				
M ellet		Patients with	SG (n=7)	46.8±2.9	6:1	42.1±4.0	3 days	NA	Factors		Histidine, proline, citrate and decanoic acid only decreased after RYGB.
2014 ³¹	Prospective conort study	obesity and T2D (N=15)	RYGB (n=8)	41.0±3.1	8:0	42.1±4.0	3 days	NA	plasma	Ontargeted GC-MS	2-hydroxybutyrate and 3-methyl-2-oxo- pentanoic acid only increase after SG.

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							1 month	45.90 ± 6.9			AAAs levels decrease induced by SG was higher. n-resol was only increased after SG
Samczuk P, 2018 ⁴⁵	Prospective cohort study	Patients with obesity and T2D (N=54)	SG (n=34)	49.3±8.7	14:20	50.92 ± 7.33	6 months	37.33 ± 7.3	Fasting serum	Untargeted GC-MS and LC-MS	Increased levels of sulfate containing Increased levels of sulfate containing metabolites were only observed after RYGB group.
			RYGB (n=20)	50.1 ± 9.3	15:5	45.79 ± 5.5	1 month	40.97 ± 5.15			PC, LPC, PE and LPE decreased after both surgeries. Sphingomyelins and choline increased after both
							6 months	32.61 ± 5.5			surgeries.
Tan G,	Prospective cohort	Patients with obesity §	SG (n=12)	36.3± 8	9:51	38.8+1.3	12 months	NA	Fasting	Targeted LC/MS (AAs and	BCAAs decreased after both surgery in similar proportions.
2016**	study	(N=22, 13 with T2D)	RYGB (n=10)	45.6±9.1			12 months	NA	serum	acylcarnitine)	AAAs levels were only significantly reduced after SG.
							3 months	46.0 ± 10.1			
			SG			C3 6 + 10 0	6 months	38.3 ±10.0			
			(n=19)			0.01 ± 0.00	9 months	33.3 ± 6.1			MAAAbalansia affaat of NVM and bishaa
							12 months	31.7 ± 4.1			wetaboloring effect of KTOB are nighter compared with SG.
							3 months	36.7 ± 5.5			BCCAs and AAAs decreased after both bariatric surgeries. Valine levels reduction was weaker
Gralka E,	Prospective cohort	Patients with	Proximal RYGB	13 6 4 1 0	CC.CT		6 months	32.6±5.4	Fasting	Untargeted ¹ H-	after SG.
2015 ²¹	study	obesity (N=106)	(n=27)			0.C I 0.C4	9 months	30.0±5.5	serum	NMR	Carboxylic acids anions levels were modified similarly by the different bariatric surgeries.
							12 months	29.4±5.9			TMAO only increased after SG.
							3 months	39.1 ± 5.5			Dimethyl sulfates increase after surgery was higher in the distal RYGR.
			Distal RYGB			46 3 + 6 5	6 months	35.2 ± 5.4			
			(n=60)				9 months	32.4±5.2			
							12 months	30.7 ± 4.9			

Table V Summary of studies comparing the metabolomic profiles induced by different bariatric surgery interventions (cont.)

$ \left \begin{array}{cccccccccccccccccccccccccccccccccccc$						RYG	RYGB with short BPL vs RYGB with long BPL	YGB with long BPL				
studyweak (u01)weak (u01)weak (u01)15.1.315.1.3mak (u01)weak (u01)mak (u01)ii	larak I,	Cross-seccional	Patients with	RYGB with short BPL (n=9)	38±3	8:1	41.8±1.1	1.6 ± 0.3 years	28.1±2.3	Fasting and post-	Untargeted ¹ H-	Fasting similar global profiles between groups
Iso is the formation	2020**	study	obesity and NGT (N=20)	RYGB with long BPL (n=11)	43±2	10:1	40.6±0.9	1.5 ± 0.3 years	26.2 ± 2.8	prandial plasma	NMR	Post-prandial levels of acetate were higher in patients submitted to RYGB with long BPL
$ \left \begin{array}{cccccccccccccccccccccccccccccccccccc$							SG vs BF	D				
Prospective colort obsity and VG (V=3) BPD (n=12) (n=3) 4.4 ± 8.2 6.6 5.1 ± 6.5 6.monts 5.atm Fasters Targeted UPC/MS Non-study (V=3) BPD (n=12) 4.4 ± 8.2 6.6 5.1 ± 6.5 5.1 ± 6.5 5.1 ± 6.5 5.1 ± 6.5 5.1 ± 6.5 5.1 ± 6.5 5.1 ± 6.5 5.1 ± 6.5 5.1 ± 6.5 1.0 ± 0.5 1.0				5G (n=25)	47.0±6.7	16:9	47.9±6.1	6 months	36.5±4.5			Lipidomic profiles induced by the two surgeries are different. BPD induced an overall decline in orbinactivide.
$\left. \begin{array}{c c c c c c c c c c c c c c c c c c c $	tamos- lolina B, 2018 ³⁹	Prospective cohort study		BPD (n=12)	44.4 ± 8.2	6:6	51.8±6.9	6 months	39.7 ± 4.4	Fasting plasma	Targeted UPLC-MS (lipidomic)	and phospholipids and an increase in bile acids levels. SG induced an increase of sphingolipids and phospholipids, and no changes in bile acids levels.
$\left \begin{array}{cccc} \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ $							BPD-DS vs I	RYGB				
Prospective cohort Prospect				BPD (n=9)	44.7±8.1	5:4	55.8±9.5	183.7 ± 61.8	39.1 ± 8.5			
BPD-DS vsADIS BPD-DS vsADIS BPD-DS vsADIS Coss-sectional study BPD-DS (n=8) 36±12 6:3 519±4.0 1.6±0.3 29.7±4.3 Fasting radiost partial Cross-sectional study BPD-DS (n=8) 36±12 6:3 51.9±4.0 1.6±0.3 29.7±4.3 Fasting radiost partial Cross-sectional study BPD-DS (n=18) SADI-S 43±7 7.2 52.0±3.7 1.5±0.3 30.0±3.6 Partial plasma	Ahlin S, 2019 ³⁸	Prospective cohort study		RYGB (n=6)	43. 7± 9.4	9:0	45.3±5.7	187.8±93.6	35.5±5.5	Fasting plasma	largeted OFLC-Wo (bile acids analysis)	No differences in fasting bile acid levels were observed between the RYGB and BPD groups.
Patients with study BPD-DS (n=9) 36±12 6:3 51.9±4.0 16±0.3 29.7±4.3 Fasting table Untargeted ¹ H- Cross-sectional study Desity and NGT (N=18) 5.01-5 5.0±3.7 15±0.3 20.7±4.3 Table Pairing Study obesity and NGT (N=18) SADI-S 43±7 7.2 52.0±3.7 1.5±0.3 30.0±3.6 plasma							BPD-DS vs S	ADI-S				
Conserectional study Obesity and MGT Obesity and MGT Integered re- number Integered re- number (N=18) SADI-S 43 ± 7 7:2 52.0 ± 3.7 1.5 ± 0.3 30.0 ± 3.6 plasma (n=9) (n=9) 43 ± 7 7:2 52.0 ± 3.7 1.5 ± 0.3 30.0 ± 3.6 plasma	reira SS,		Patients with	(0=u)	36±12	6:3	51.9±4.0	1.6±0.3 years	29.7 ± 4.3	Fasting	Li bostonostel I	Fasting similar global profiles between groups
	202055	study	obesity and NGT (N= 18)	S-IDAS (n=9)	43±7	7:2	52.0±3.7	1.5 ± 0.3 years	30.0±3.6	prandial plasma	Undigered 1- NMR	Post-prandial levels of BCAAs were higher in SADI-5 group

Table V Summary of studies comparing the metabolomic profiles induced by different bariatric surgery interventions (cont.)

Gas chromatography: ¹H-NIMR - Proton nuclear magnetic resonance; LAGB - Laparoscopic adjustable gastric band; . LC. Liquid chromatography, MS - Mass Spectrometry, NA - Not available; NGT- Normal glucose tolerance; . PC - Phosphatidylcholines, LP - Biopancreatic investor with Gastric Bypass; SADI-S- Single Anastomosis with duodeno-fleal bypass with seve gastrectomy; 7720 - Type 2 dabetes; TMAO - Trimethylamine N-cxide; UPC. Ultra-Performance Liquid Chromatography

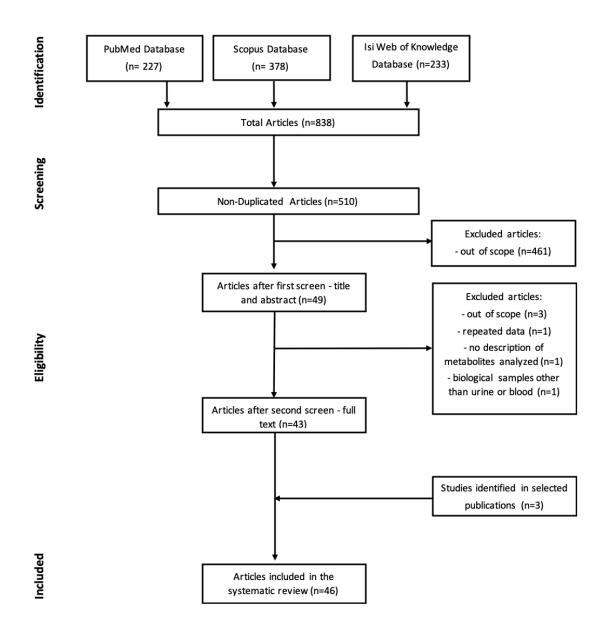
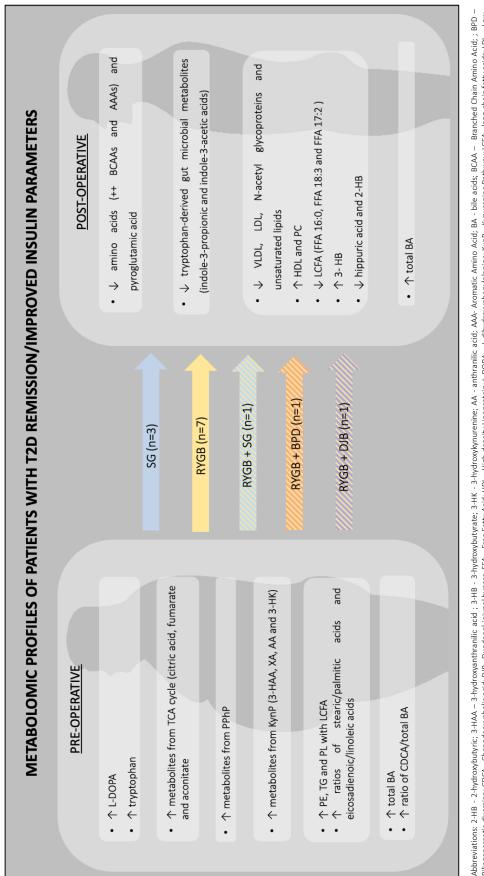
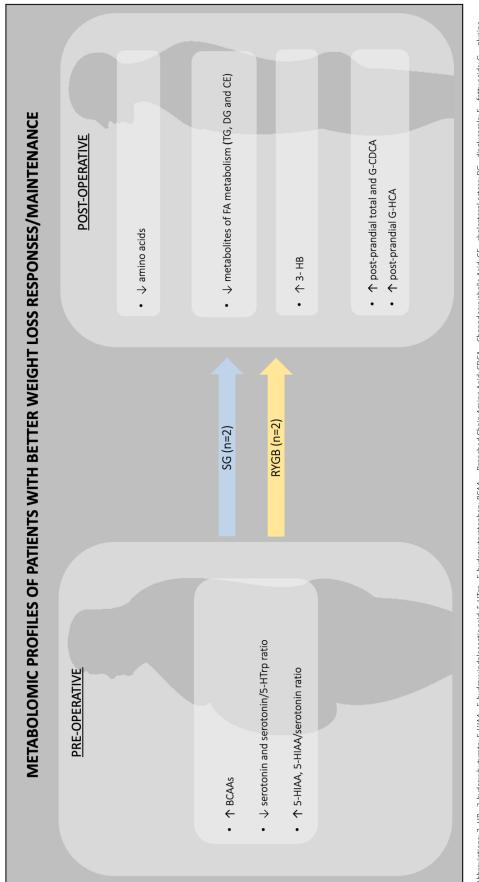


Figure 1 Flowchart of the search, eligibility criteria approaches and study inclusion for systematic review



Abbreviations: 2-HB - 2-hydroxybutyric; 3-HAA - 3-hydroxyanthranilic acid ; 3-HB - 3-hydroxybutyrate; 3-HK - 3-hydroxybutyrate; 3-HY - 3-hydroxybutyrate; 3-HK - 3-hydroxypate; 3-HK - 3-HY - 3-hydroxypate; 3-HK - 3-hydroxypate; 3-HK - 3-hydroxypate; 3-HK - 3-HK

Figure 2 Metabolomic profiles of patients with T2D remission/improved insulin parameters



Abbreviations: 3-HB - 3-hydroxybutyrate; 5-HIA - 5-hydroxyindoleacetic acid; 5-HTrp - 5-hydroxytryptophan; BCAA – Branched Chain Amino Acid; CDCA – Chenodeoxycholic Acid; CE - cholesterol esters; DG - diaglycerols; F – fatty acids; G-- glycine amidated; HCA - hyocholic acid; SG – Sleeve gastrectomy; RYGB - Roux-en-Y Gastric Bypass; TG - triacylglycerols

Figure 3 Metabolomic profiles of patients with better weight loss responses/maintenance

References | Bibliografia

- 1. Rangel-Huerta OD, Pastor-Villaescusa B, Gil A. Are we close to defining a metabolomic signature of human obesity? A systematic review of metabolomics studies. Metabolomics. 2019;15(6):93.
- 2. Nyberg ST, Batty GD, Pentti J, Virtanen M, Alfredsson L, Fransson EI, Goldberg M, Heikkilä K, Jokela M, Knutsson A, Koskenvuo M, Lallukka T, Leineweber C, Lindbohm JV, Madsen IEH, Magnusson Hanson LL, Nordin M, Oksanen T, Pietiläinen O, Rahkonen O, Rugulies R, Shipley MJ, Stenholm S, Suominen S, Theorell T, Vahtera J, Westerholm PJM, Westerlund H, Zins M, Hamer M, Singh-Manoux A, Bell JA, Ferrie JE, Kivimäki M. Obesity and loss of disease-free years owing to major non-communicable diseases: a multicohort study. The Lancet Public Health. 2018;3(10):e490-e497.
- Tulipani S, Griffin J, Palau-Rodriguez M, Mora-Cubillos X, Bernal-Lopez RM, Tinahones FJ, Corkey BE, Andres-Lacueva C. Metabolomics-guided insights on bariatric surgery versus behavioral interventions for weight loss. Obesity (Silver Spring). 2016;24(12):2451-2466.
- 4. Poirier P, Cornier MA, Mazzone T, Stiles S, Cummings S, Klein S, McCullough PA, Ren Fielding C, Franklin BA. Bariatric surgery and cardiovascular risk factors: a scientific statement from the American Heart Association. Circulation. 2011;123(15):1683-1701.
- 5. Samczuk P, Ciborowski M, Kretowski A. Application of Metabolomics to Study Effects of Bariatric Surgery. J Diabetes Res. 2018;2018:6270875.
- 6. Luo JN, Sheu EG. Do Serum Metabolites Predict Weight Regain Following Bariatric Surgery? Dig Dis Sci. 2020;65(4):922-924.
- Peterli R, Steinert RE, Woelnerhanssen B, Peters T, Christoffel-Courtin C, Gass M, Kern B, von Fluee M, Beglinger C. Metabolic and hormonal changes after laparoscopic Roux-en-Y gastric bypass and sleeve gastrectomy: a randomized, prospective trial. Obes Surg. 2012;22(5):740-748.
- 8. Floegel A, Stefan N, Yu Z, Mühlenbruch K, Drogan D, Joost HG, Fritsche A, Häring HU, Hrabě de Angelis M, Peters A, Roden M, Prehn C, Wang-Sattler R, Illig T, Schulze MB, Adamski J, Boeing H, Pischon T. Identification of serum metabolites associated with risk of type 2 diabetes using a targeted metabolomic approach. Diabetes. 2013;62(2):639-648.
- 9. Malin SK, Kashyap SR. Effects of various gastrointestinal procedures on β -cell function in obesity and type 2 diabetes. Surg Obes Relat Dis. 2016;12(6):1213-1219.
- Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, Shamseer L, Tetzlaff JM, Akl EA, Brennan SE, Chou R, Glanville J, Grimshaw JM, Hróbjartsson A, Lalu MM, Li T, Loder EW, Mayo-Wilson E, McDonald S, McGuinness LA, Stewart LA, Thomas J, Tricco AC, Welch VA, Whiting P, Moher D. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. PLoS Med. 2021;18(3):e1003583.
- 11. Luo P, Yu H, Zhao X, Bao Y, Hong CS, Zhang P, Tu Y, Yin P, Gao P, Wei L, Zhuang Z, Jia W, Xu G. Metabolomics Study of Roux-en-Y Gastric Bypass Surgery (RYGB) to Treat Type 2 Diabetes Patients Based on Ultraperformance Liquid Chromatography-Mass Spectrometry. J Proteome Res. 2016;15(4):1288-1299.
- 12. Magkos F, Bradley D, Schweitzer GG, Finck BN, Eagon JC, Ilkayeva O, Newgard CB, Klein S. Effect of Roux-en-Y gastric bypass and laparoscopic adjustable gastric banding on branched-chain amino acid metabolism. Diabetes. 2013;62(8):2757-2761.
- 13. Mutch DM, Fuhrmann JC, Rein D, Wiemer JC, Bouillot JL, Poitou C, Clément K. Metabolite profiling identifies candidate markers reflecting the clinical adaptations associated with Roux-en-Y gastric bypass surgery. PLoS One. 2009;4(11):e7905.
- 14. Narath SH, Mautner SI, Svehlikova E, Schultes B, Pieber TR, Sinner FM, Gander E, Libiseller G, Schimek MG, Sourij H, Magnes C. An Untargeted Metabolomics Approach to

Characterize Short-Term and Long-Term Metabolic Changes after Bariatric Surgery. PLoS One. 2016;11(9):e0161425.

- 15. Palau-Rodriguez M, Tulipani S, Marco-Ramell A, Miñarro A, Jáuregui O, Sanchez-Pla A, Ramos-Molina B, Tinahones FJ, Andres-Lacueva C. Metabotypes of response to bariatric surgery independent of the magnitude of weight loss. PLoS One. 2018;13(6):e0198214.
- 16. Wijayatunga NN, Sams VG, Dawson JA, Mancini ML, Mancini GJ, Moustaid-Moussa N. Roux-en-Y gastric bypass surgery alters serum metabolites and fatty acids in patients with morbid obesity. Diabetes Metab Res Rev. 2018;34(8):e3045.
- 17. Yao J, Kovalik JP, Lai OF, Lee PC, Eng A, Chan WH, Tham KW, Lim E, Bee YM, Tan HC. Comprehensive Assessment of the Effects of Sleeve Gastrectomy on Glucose, Lipid, and Amino Acid Metabolism in Asian Individuals with Morbid Obesity. Obes Surg. 2019;29(1):149-158.
- 18. Yoshida N, Kitahama S, Yamashita T, Hirono Y, Tabata T, Saito Y, Shinohara R, Nakashima H, Emoto T, Hirota Y, Takahashi T, Ogawa W, Hirata KI. Metabolic alterations in plasma after laparoscopic sleeve gastrectomy. J Diabetes Investig. 2021;12(1):123-129.
- 19. Arora T, Velagapudi V, Pournaras DJ, Welbourn R, le Roux CW, Orešič M, Bäckhed F. Rouxen-Y Gastric Bypass Surgery Induces Early Plasma Metabolomic and Lipidomic Alterations in Humans Associated with Diabetes Remission. PLoS One. 2015;10(5):e0126401.
- 20. Dadson P, Rebelos E, Honka H, Juárez-Orozco LE, Kalliokoski KK, Iozzo P, Teuho J, Salminen P, Pihlajamäki J, Hannukainen JC, Nuutila P. Change in abdominal, but not femoral subcutaneous fat CT-radiodensity is associated with improved metabolic profile after bariatric surgery. Nutr Metab Cardiovasc Dis. 2020;30(12):2363-2371.
- 21. Gralka E, Luchinat C, Tenori L, Ernst B, Thurnheer M, Schultes B. Metabolomic fingerprint of severe obesity is dynamically affected by bariatric surgery in a procedure-dependent manner. Am J Clin Nutr. 2015;102(6):1313-1322.
- 22. Khoo CM, Muehlbauer MJ, Stevens RD, Pamuklar Z, Chen J, Newgard CB, Torquati A. Postprandial metabolite profiles reveal differential nutrient handling after bariatric surgery compared with matched caloric restriction. Ann Surg. 2014;259(4):687-693.
- 23. Kwon Y, Jang M, Lee Y, Ha J, Park S. Metabolomic Analysis of the Improvements in Insulin Secretion and Resistance After Sleeve Gastrectomy: Implications of the Novel Biomarkers. Obes Surg. 2021;31(1):43-52.
- 24. Laferrère B, Reilly D, Arias S, Swerdlow N, Gorroochurn P, Bawa B, Bose M, Teixeira J, Stevens RD, Wenner BR, Bain JR, Muehlbauer MJ, Haqq A, Lien L, Shah SH, Svetkey LP, Newgard CB. Differential metabolic impact of gastric bypass surgery versus dietary intervention in obese diabetic subjects despite identical weight loss. Sci Transl Med. 2011;3(80):80re82.
- 25. Lips MA, Van Klinken JB, van Harmelen V, Dharuri HK, t Hoen PA, Laros JF, van Ommen GJ, Janssen IM, Van Ramshorst B, Van Wagensveld BA, Swank DJ, Van Dielen F, Dane A, Harms A, Vreeken R, Hankemeier T, Smit JW, Pijl H, Willems van Dijk K. Roux-en-Y gastric bypass surgery, but not calorie restriction, reduces plasma branched-chain amino acids in obese women independent of weight loss or the presence of type 2 diabetes. Diabetes Care. 2014;37(12):3150-3156.
- 26. Lopes TI, Geloneze B, Pareja JC, Calixto AR, Ferreira MM, Marsaioli AJ. Blood Metabolome Changes Before and After Bariatric Surgery: A (1)H NMR-Based Clinical Investigation. Omics. 2015;19(5):318-327.
- 27. Cabré N, Luciano-Mateo F, Baiges-Gayà G, Fernández-Arroyo S, Rodríguez-Tomàs E, Hernández-Aguilera A, París M, Sabench F, Del Castillo D, López-Miranda J, Menéndez JA, Camps J, Joven J. Plasma metabolic alterations in patients with severe obesity and nonalcoholic steatohepatitis. Aliment Pharmacol Ther. 2020;51(3):374-387.
- 28. Hubal MJ, Nadler EP, Ferrante SC, Barberio MD, Suh JH, Wang J, Dohm GL, Pories WJ, Mietus-Snyder M, Freishtat RJ. Circulating Adipocyte-Derived Exosomal MicroRNAs

Associated with Decreased Insulin Resistance After Gastric Bypass. Obesity. 2017;25(1):102-110.

- 29. Samczuk P, Luba M, Godzien J, Mastrangelo A, Hady HR, Dadan J, Barbas C, Gorska M, Kretowski A, Ciborowski M. "Gear mechanism" of bariatric interventions revealed by untargeted metabolomics. J Pharm Biomed Anal. 2018;151:219-226.
- 30. Mendonça Machado N, Torrinhas RS, Sala P, Ishida RK, Guarda I, Moura EGH, Sakai P, Santo MA, Linetzky Waitzberg D. Type 2 Diabetes Metabolic Improvement After Roux-en-Y Gastric Bypass May Include a Compensatory Mechanism That Balances Fatty Acid β and ω Oxidation. JPEN J Parenter Enteral Nutr. 2020;44(8):1417-1427.
- 31. Jüllig M, Yip S, Xu A, Smith G, Middleditch M, Booth M, Babor R, Beban G, Murphy R. Lower fetuin-A, retinol binding protein 4 and several metabolites after gastric bypass compared to sleeve gastrectomy in patients with type 2 diabetes. PLoS One. 2014;9(5):e96489.
- 32. Ocaña-Wilhelmi L, Cardona F, Garrido-Sanchez L, Fernandez-Garcia D, Tinahones FJ, Ramos-Molina B. Change in serum polyamine metabolome pattern after bariatric surgery in obese patients with metabolic syndrome. Surg Obes Relat Dis. 2020;16(2):306-311.
- Sarosiek K, Pappan KL, Gandhi AV, Saxena S, Kang CY, McMahon H, Chipitsyna GI, Tichansky DS, Arafat HA. Conserved Metabolic Changes in Nondiabetic and Type 2 Diabetic Bariatric Surgery Patients: Global Metabolomic Pilot Study. J Diabetes Res. 2016;2016:3467403.
- 34. Oberbach A, Blüher M, Wirth H, Till H, Kovacs P, Kullnick Y, Schlichting N, Tomm JM, Rolle-Kampczyk U, Murugaiyan J, Binder H, Dietrich A, von Bergen M. Combined proteomic and metabolomic profiling of serum reveals association of the complement system with obesity and identifies novel markers of body fat mass changes. J Proteome Res. 2011;10(10):4769-4788.
- Herzog K, Berggren J, Al Majdoub M, Balderas Arroyo C, Lindqvist A, Hedenbro J, Groop L, Wierup N, Spégel P. Metabolic Effects of Gastric Bypass Surgery: Is It All About Calories? Diabetes. 2020;69(9):2027-2035.
- 36. Fiamoncini J, Fernandes Barbosa C, Arnoni Junior JR, Araújo Junior JC, Taglieri C, Szego T, Gelhaus B, Possolo de Souza H, Daniel H, Martins de Lima T. Roux-en-Y Gastric Bypass Surgery Induces Distinct but Frequently Transient Effects on Acylcarnitine, Bile Acid and Phospholipid Levels. Metabolites. 2018;8(4).
- 37. Mika A, Kaczynski Z, Stepnowski P, Kaczor M, Proczko-Stepaniak M, Kaska L, Sledzinski T. Potential Application of H-1 NMR for Routine Serum Lipidome Analysis - Evaluation of Effects of Bariatric Surgery. Scientific Reports. 2017;7.
- Ahlin S, Cefalù C, Bondia-Pons I, Capristo E, Marini L, Gastaldelli A, Mingrone G, Nolan JJ.
 Bile acid changes after metabolic surgery are linked to improvement in insulin sensitivity.
 British Journal of Surgery. 2019;106(9):1178-1186.
- Ramos-Molina B, Castellano-Castillo D, Alcaide-Torres J, Pastor Ó, de Luna Díaz R, Salas-Salvadó J, López-Moreno J, Fernández-García JC, Macías-González M, Cardona F, Tinahones FJ. Differential effects of restrictive and malabsorptive bariatric surgery procedures on the serum lipidome in obese subjects. J Clin Lipidol. 2018;12(6):1502-1512.
- 40. Ahmad NN, Pfalzer A, Kaplan LM. Roux-en-Y gastric bypass normalizes the blunted postprandial bile acid excursion associated with obesity. International Journal of Obesity. 2013;37(12):1553-1559.
- 41. Kindel TL, Krause C, Helm MC, McBride CL, Oleynikov D, Thakare R, Alamoudi J, Kothari V, Alnouti Y, Kohli R. Increased glycine-amidated hyocholic acid correlates to improved early weight loss after sleeve gastrectomy. Surgical Endoscopy and Other Interventional Techniques. 2018;32(2):805-812.
- 42. Kayser BD, Lhomme M, Dao MC, Ichou F, Bouillot JL, Prifti E, Kontush A, Chevallier JM, Aron-Wisnewsky J, Dugail I, Clément K. Serum lipidomics reveals early differential effects

of gastric bypass compared with banding on phospholipids and sphingolipids independent of differences in weight loss. Int J Obes (Lond). 2017;41(6):917-925.

- 43. Friedrich N, Budde K, Wolf T, Jungnickel A, Grotevendt A, Dressler M, Völzke H, Blüher M, Nauck M, Lohmann T, Wallaschofksi H. Short-term changes of the urine metabolome after bariatric surgery. Omics. 2012;16(11):612-620.
- 44. Shantavasinkul PC, Muehlbauer MJ, Bain JR, Ilkayeva OR, Craig DM, Newgard CB, Svetkey LP, Shah SH, Torquati A. Improvement in insulin resistance after gastric bypass surgery is correlated with a decline in plasma 2-hydroxybutyric acid. Surg Obes Relat Dis. 2018;14(8):1126-1132.
- 45. Samczuk P, Hady HR, Adamska-Patruno E, Citko A, Dadan J, Barbas C, Kretowski A, Ciborowski M. In-and-Out Molecular Changes Linked to the Type 2 Diabetes Remission after Bariatric Surgery: An Influence of Gut Microbes on Mitochondria Metabolism. Int J Mol Sci. 2018;19(12).
- 46. Tan HC, Khoo CM, Tan MZ, Kovalik JP, Ng AC, Eng AK, Lai OF, Ching JH, Tham KW, Pasupathy S. The Effects of Sleeve Gastrectomy and Gastric Bypass on Branched-Chain Amino Acid Metabolism 1 Year After Bariatric Surgery. Obes Surg. 2016;26(8):1830-1835.
- 47. Lopes TI, Geloneze B, Pareja JC, Calixto AR, Ferreira MM, Marsaioli AJ. "Omics" Prospective Monitoring of Bariatric Surgery: Roux-En-Y Gastric Bypass Outcomes Using Mixed-Meal Tolerance Test and Time-Resolved (1)H NMR-Based Metabolomics. Omics. 2016;20(7):415-423.
- 48. Yu H, Ni Y, Bao Y, Zhang P, Zhao A, Chen T, Xie G, Tu Y, Zhang L, Su M, Wei L, Jia W, Jia W. Chenodeoxycholic Acid as a Potential Prognostic Marker for Roux-en-Y Gastric Bypass in Chinese Obese Patients. J Clin Endocrinol Metab. 2015;100(11):4222-4230.
- 49. Zhao L, Ni Y, Yu H, Zhang P, Zhao A, Bao Y, Liu J, Chen T, Xie G, Panee J, Chen W, Rajani C, Wei R, Su M, Jia W, Jia W. Serum stearic acid/palmitic acid ratio as a potential predictor of diabetes remission after Roux-en-Y gastric bypass in obesity. Faseb j. 2017;31(4):1449-1460.
- 50. Li QR, Wang ZM, Albrechtsen NJW, Wang DD, Su ZD, Gao XF, Wu QQ, Zhang HP, Zhu L, Li RX, Jacobsen S, Jorgensen NB, Dirksen C, Bojsen-Moller KN, Petersen JS, Madsbad S, Clausen TR, Diderichsen B, Chen LN, Holst JJ, Zeng R, Wu JR. Systems Signatures Reveal Unique Remission-path of Type 2 Diabetes Following Roux-en-Y Gastric Bypass Surgery. Ebiomedicine. 2018;28:234-240.
- 51. Ha J, Jang M, Kwon Y, Park YS, Park DJ, Lee JH, Lee HJ, Ha TK, Kim YJ, Han SM, Han SU, Heo Y, Park S. Metabolomic Profiles Predict Diabetes Remission after Bariatric Surgery. J Clin Med. 2020;9(12).
- 52. Kwon HN, Lee YJ, Kang JH, Choi JH, An YJ, Kang S, Lee DH, Suh YJ, Heo Y, Park S. Prediction of glycated hemoglobin levels at 3 months after metabolic surgery based on the 7-day plasma metabolic profile. PLoS One. 2014;9(11):e109609.
- 53. Kwon Y, Jang M, Lee Y, Ha J, Park S. Amino Acid Metabolites and Slow Weight Loss in the Early Postoperative Period after Sleeve Gastrectomy. J Clin Med. 2020;9(8).
- 54. Abidi W, Nestoridi E, Feldman H, Stefater M, Clish C, Thompson CC, Stylopoulos N. Differential Metabolomic Signatures in Patients with Weight Regain and Sustained Weight Loss After Gastric Bypass Surgery: A Pilot Study. Digestive Diseases and Sciences. 2020;65(4):1144-1154.
- 55. Pereira SS, Jarak I, Carvalho RA, Oliveira PF, Alves MG, Guimarães M, Almeida R, Pereira AM, Wewer Albrechtsen NJ, Holst JJ, Nora M, Monteiro MP. Different Malabsorptive Obesity Surgery Interventions Result in Distinct Postprandial Amino Acid Metabolomic Signatures. Obes Surg. 2020;30(10):4019-4028.
- 56. Jarak I, Pereira SS, Carvalho RA, Oliveira PF, Alves MG, Guimarães M, Wewer Albrechtsen NJ, Holst JJ, Nora M, Monteiro MP. Gastric Bypass with Different Biliopancreatic Limb Lengths Results in Similar Post-absorptive Metabolomics Profiles. Obes Surg. 2020;30(3):1068-1078.

- 57. Rauschert S, Uhl O, Koletzko B, Hellmuth C. Metabolomic biomarkers for obesity in humans: a short review. Ann Nutr Metab. 2014;64(3-4):314-324.
- 58. Newgard CB, An J, Bain JR, Muehlbauer MJ, Stevens RD, Lien LF, Haqq AM, Shah SH, Arlotto M, Slentz CA, Rochon J, Gallup D, Ilkayeva O, Wenner BR, Yancy WS, Jr., Eisenson H, Musante G, Surwit RS, Millington DS, Butler MD, Svetkey LP. A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and contributes to insulin resistance. Cell Metab. 2009;9(4):311-326.
- 59. Yabut JM, Crane JD, Green AE, Keating DJ, Khan WI, Steinberg GR. Emerging Roles for Serotonin in Regulating Metabolism: New Implications for an Ancient Molecule. Endocr Rev. 2019;40(4):1092-1107.
- 60. She P, Van Horn C, Reid T, Hutson SM, Cooney RN, Lynch CJ. Obesity-related elevations in plasma leucine are associated with alterations in enzymes involved in branched-chain amino acid metabolism. Am J Physiol Endocrinol Metab. 2007;293(6):E1552-1563.
- 61. Guasch-Ferré M, Hruby A, Toledo E, Clish CB, Martínez-González MA, Salas-Salvadó J, Hu FB. Metabolomics in Prediabetes and Diabetes: A Systematic Review and Meta-analysis. Diabetes Care. 2016;39(5):833-846.
- 62. Felig P, Marliss E, Cahill GF, Jr. Plasma amino acid levels and insulin secretion in obesity. N Engl J Med. 1969;281(15):811-816.
- 63. Alves A, Bassot A, Bulteau A-L, Pirola L, Morio B. Glycine Metabolism and Its Alterations in Obesity and Metabolic Diseases. Nutrients. 2019;11(6):1356.
- 64. Wang-Sattler R, Yu Z, Herder C, Messias AC, Floegel A, He Y, Heim K, Campillos M, Holzapfel C, Thorand B, Grallert H, Xu T, Bader E, Huth C, Mittelstrass K, Döring A, Meisinger C, Gieger C, Prehn C, Roemisch-Margl W, Carstensen M, Xie L, Yamanaka-Okumura H, Xing G, Ceglarek U, Thiery J, Giani G, Lickert H, Lin X, Li Y, Boeing H, Joost HG, de Angelis MH, Rathmann W, Suhre K, Prokisch H, Peters A, Meitinger T, Roden M, Wichmann HE, Pischon T, Adamski J, Illig T. Novel biomarkers for pre-diabetes identified by metabolomics. Mol Syst Biol. 2012;8:615.
- 65. Caldow MK, Ham DJ, Godeassi DP, Chee A, Lynch GS, Koopman R. Glycine supplementation during calorie restriction accelerates fat loss and protects against further muscle loss in obese mice. Clin Nutr. 2016;35(5):1118-1126.
- 66. Rowland I, Gibson G, Heinken A, Scott K, Swann J, Thiele I, Tuohy K. Gut microbiota functions: metabolism of nutrients and other food components. Eur J Nutr. 2018;57(1):1-24.
- 67. Magne F, Gotteland M, Gauthier L, Zazueta A, Pesoa S, Navarrete P, Balamurugan R. The Firmicutes/Bacteroidetes Ratio: A Relevant Marker of Gut Dysbiosis in Obese Patients? Nutrients. 2020;12(5).
- 68. Smith EA, Macfarlane GT. Enumeration of human colonic bacteria producing phenolic and indolic compounds: effects of pH, carbohydrate availability and retention time on dissimilatory aromatic amino acid metabolism. J Appl Bacteriol. 1996;81(3):288-302.
- 69. Zhang YJ, Li S, Gan RY, Zhou T, Xu DP, Li HB. Impacts of gut bacteria on human health and diseases. Int J Mol Sci. 2015;16(4):7493-7519.
- 70. Murphy R, Tsai P, Jüllig M, Liu A, Plank L, Booth M. Differential Changes in Gut Microbiota After Gastric Bypass and Sleeve Gastrectomy Bariatric Surgery Vary According to Diabetes Remission. Obesity Surgery. 2017;27(4):917-925.
- 71. Ohland CL, Jobin C. Microbial activities and intestinal homeostasis: A delicate balance between health and disease. Cell Mol Gastroenterol Hepatol. 2015;1(1):28-40.
- 72. Pereira PR, Guimarães M, Morais T, Pereira SS, Nora M, Monteiro MP. Diabetic and Elder Patients Experience Superior Cardiovascular Benefits After Gastric Bypass Induced Weight Loss. Front Endocrinol (Lausanne). 2018;9:718.
- 73. Ammar W, Basset HA, Al Faramawy A, Hegazy T, Sharaf Y. Bariatric surgery and cardiovascular outcome. The Egyptian Heart Journal. 2020;72(1):67.

- 74. Trøseid M, Hov JR, Nestvold TK, Thoresen H, Berge RK, Svardal A, Lappegård KT. Major Increase in Microbiota-Dependent Proatherogenic Metabolite TMAO One Year After Bariatric Surgery. Metabolic syndrome and related disorders. 2016;14(4):197-201.
- 75. He X, Slupsky CM. Metabolic Fingerprint of Dimethyl Sulfone (DMSO2) in Microbial– Mammalian Co-metabolism. Journal of Proteome Research. 2014;13(12):5281-5292.
- 76. Patel DP, Krausz KW, Xie C, Beyoğlu D, Gonzalez FJ, Idle JR. Metabolic profiling by gas chromatography-mass spectrometry of energy metabolism in high-fat diet-fed obese mice. PLoS One. 2017;12(5):e0177953.
- 77. Cotter DG, Schugar RC, Crawford PA. Ketone body metabolism and cardiovascular disease. Am J Physiol Heart Circ Physiol. 2013;304(8):H1060-1076.
- 78. Park S, Kim DS, Daily JW. Central infusion of ketone bodies modulates body weight and hepatic insulin sensitivity by modifying hypothalamic leptin and insulin signaling pathways in type 2 diabetic rats. Brain Res. 2011;1401:95-103.
- Li JV, Ashrafian H, Bueter M, Kinross J, Sands C, le Roux CW, Bloom SR, Darzi A, Athanasiou T, Marchesi JR, Nicholson JK, Holmes E. Metabolic surgery profoundly influences gut microbial-host metabolic cross-talk. Gut. 2011;60(9):1214-1223.
- Goncalves D, Barataud A, De Vadder F, Vinera J, Zitoun C, Duchampt A, Mithieux G. Bile Routing Modification Reproduces Key Features of Gastric Bypass in Rat. Ann Surg. 2015;262(6):1006-1015.
- 81. Thomas C, Gioiello A, Noriega L, Strehle A, Oury J, Rizzo G, Macchiarulo A, Yamamoto H, Mataki C, Pruzanski M, Pellicciari R, Auwerx J, Schoonjans K. TGR5-mediated bile acid sensing controls glucose homeostasis. Cell Metab. 2009;10(3):167-177.
- 82. Russell DW. Fifty years of advances in bile acid synthesis and metabolism. J Lipid Res. 2009;50 Suppl(Suppl):S120-125.
- 83. Watanabe M, Houten SM, Mataki C, Christoffolete MA, Kim BW, Sato H, Messaddeq N, Harney JW, Ezaki O, Kodama T, Schoonjans K, Bianco AC, Auwerx J. Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. Nature. 2006;439(7075):484-489.
- 84. Nagao K, Yanagita T. Medium-chain fatty acids: functional lipids for the prevention and treatment of the metabolic syndrome. Pharmacol Res. 2010;61(3):208-212.
- 85. Han JR, Deng B, Sun J, Chen CG, Corkey BE, Kirkland JL, Ma J, Guo W. Effects of dietary medium-chain triglyceride on weight loss and insulin sensitivity in a group of moderately overweight free-living type 2 diabetic Chinese subjects. Metabolism. 2007;56(7):985-991.
- 86. Wein S, Wolffram S, Schrezenmeir J, Gasperiková D, Klimes I, Seböková E. Medium-chain fatty acids ameliorate insulin resistance caused by high-fat diets in rats. Diabetes Metab Res Rev. 2009;25(2):185-194.
- 87. Poisson J-PG, Cunnane SC. Long-chain fatty acid metabolism in fasting and diabetes: relation between altered desaturase activity and fatty acid composition. The Journal of Nutritional Biochemistry. 1991;2(2):60-70.
- 88. Lemaitre RN, Fretts AM, Sitlani CM, Biggs ML, Mukamal K, King IB, Song X, Djoussé L, Siscovick DS, McKnight B, Sotoodehnia N, Kizer JR, Mozaffarian D. Plasma phospholipid very-long-chain saturated fatty acids and incident diabetes in older adults: the Cardiovascular Health Study. Am J Clin Nutr. 2015;101(5):1047-1054.
- 89. Rhee EP, Cheng S, Larson MG, Walford GA, Lewis GD, McCabe E, Yang E, Farrell L, Fox CS, O'Donnell CJ, Carr SA, Vasan RS, Florez JC, Clish CB, Wang TJ, Gerszten RE. Lipid profiling identifies a triacylglycerol signature of insulin resistance and improves diabetes prediction in humans. J Clin Invest. 2011;121(4):1402-1411.
- 90. Liang H, Tantiwong P, Sriwijitkamol A, Shanmugasundaram K, Mohan S, Espinoza S, Defronzo RA, Dubé JJ, Musi N. Effect of a sustained reduction in plasma free fatty acid concentration on insulin signalling and inflammation in skeletal muscle from human subjects. J Physiol. 2013;591(11):2897-2909.

- 91. Rauschert S, Uhl O, Koletzko B, Kirchberg F, Mori TA, Huang R-C, Beilin LJ, Hellmuth C, Oddy WH. Lipidomics Reveals Associations of Phospholipids With Obesity and Insulin Resistance in Young Adults. The Journal of Clinical Endocrinology & Metabolism. 2016;101(3):871-879.
- 92. Monzo-Beltran L, Vazquez-Tarragón A, Cerdà C, Garcia-Perez P, Iradi A, Sánchez C, Climent B, Tormos C, Vázquez-Prado A, Girbés J, Estáñ N, Blesa S, Cortés R, Chaves FJ, Sáez GT. One-year follow-up of clinical, metabolic and oxidative stress profile of morbid obese patients after laparoscopic sleeve gastrectomy. 8-oxo-dG as a clinical marker. Redox Biology. 2017;12:389-402.
- 93. Iqbal J, Walsh MT, Hammad SM, Hussain MM. Sphingolipids and Lipoproteins in Health and Metabolic Disorders. Trends in Endocrinology & Metabolism. 2017;28(7):506-518.

Supplementary File 1

DATABASE SEARCH

PUBMED

("bariatric surger*" OR "bariatric intervention*" OR "obesity surger*" OR "bariatric procedur*"OR "gastric bypass" OR "biliopancreatic diversion" OR "RYGB" OR "sleeve gastrectomy" OR "BPD" OR "BPD-DS" OR "duodenal switch" OR "Single anastomosis duodenoileal bypass" OR "SADI-S" OR "SADI" OR "SIPS") AND (metabolomic* OR metabonomic* OR metabolite* OR "metabolome") NOT ("bronchopulmonary dysplasia" OR infant* OR child* OR "gestational" OR pregnanc* OR animal* OR "rat" OR "rats" OR "mice" OR "mouse")

<u>SCOPUS</u>

TITLE-ABS-KEY ("bariatric surger*" OR "bariatric intervention*" OR "obesity surger*" OR "bariatric procedur*"OR "gastric bypass" OR "biliopancreatic diversion" OR "RYGB" OR "sleeve gastrectomy" OR "BPD" OR "BPD-DS" OR "duodenal switch" OR "Single anastomosis duodenoileal bypass" OR "SADI-S" OR "SADI" OR "SIPS") AND TITLE-ABS-KEY (metabolomic* OR metabonomic* OR metabolite* OR "metabolome") AND NOT TITLE-ABS-KEY ("bronchopulmonary dysplasia" OR "Borderline personality disorder" OR infant* OR child* OR "gestational" OR pregnanc* OR animal* OR "rat" OR "rats" OR "mice" OR "mouse")

WEB OF SCIENCE

#1: ((TS=("bariatric surger*" OR "bariatric intervention*" OR "obesity surger*" OR "bariatric procedur*"OR "gastric bypass" OR "biliopancreatic diversion" OR "RYGB" OR "sleeve gastrectomy" OR "BPD" OR "BPD-DS" OR "duodenal switch" OR "Single anastomosis duodenoileal bypass" OR "SADI-S" OR "SADI" OR "SIPS")) NOT (TS=("bronchopulmonary dysplasia " OR "Borderline personality disorder" OR "infants" OR "pregnancy" OR "children" OR "animal" OR "gestational"))) AND LANGUAGE: (English) AND DOCUMENT TYPES: (Article)

Indexes = SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH, ESCI, CCR-EXPANDED, IC Timespan=All years

#2: ((TS=(metabolomic* OR metabonomic* OR metabolite* OR metabolome))) AND LANGUAGE: (English) AND DOCUMENT TYPES: (Article)

Indexes=SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH, ESCI, CCR-EXPANDED, IC Timespan=All years

#3 Final Search: #1 AND #2