

P2334 Identification of a conserved operon in chlorhexidine tolerant *Enterococcus faecium* from different clades and origins

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Background: Chlorhexidine-gluconate (CHX) activity against *Enterococcus faecium*-*Efm* is scarcely documented, with most available data not addressing the clonal background of the strains (clades A1-infection derived strains, A2-mostly animals, B-human commensal). A P102H-mutation in a conserved DNA-binding-response-regulator (ChtR) has been associated with chlorhexidine tolerance among strains of *Efm* clade A1, although the regulon remained unidentified (PMID:28242664). Here, we evaluated CHX activity, the distribution of ChtR-P102H, the predicted ChtR regulon and its variability among *Efm* from diverse sources and clades.

Materials/methods: *Efm* (n=106) from clades A1 (n=48; human/animal/food/environment), A2 (n=43; human/animal/food) and B (n=15; human/animal/environment) (1995-2016; 5-countries; multidrug-resistant:72%) were included. CHX susceptibility (range:2-32mg/L) was determined by broth-microdilution. *Efm* MIC distribution was analysed by ECOFFINDER-tool (http://www.eucast.org/mic_distributions_and_ecoffs/). Thirty-five *Efm* were sequenced (Illumina-NextSeq platform/2X150bp paired-end). DOOR software (<http://csbl.bmb.uga.edu/DOOR/index.php>) predicted ChtR regulon. Amino-acid mutations in ChtR and other operon proteins were identified by comparison (BLASTp-NCBI) with the CHX-tolerant reference strain ChtR-P102H-*Efm*-E1162 (EFF34003.1; PMID:28242664).

Results: CHX-MIC ranged between ≤ 2 -32mg/L (mode=16mg/L; 51% of isolates), with the MICs fitted curve slightly deviated to the left comparing to raw data distribution, suggesting the presence of a non-wild-*Efm* population. Most of *Efm* with a MIC ≥ 8 mg/L (96%-n=25/26; 3 clades; 14 of infections) presented the ChtR-P102H, while most isolates with a MIC ≤ 4 mg/L did not (67%-n=6/9; clades A2/B; 2 of infections). The predicted 4101bp-operon associated with *chtR* included a previously identified sensor-histidine-kinase as well as a DMT superfamily drug/metabolite transporter and an amino-acid-polyamine-organocation family transporter genes, firstly described here. The complete operon was present in all 35 *Efm* sequenced. The 25 *Efm*-MIC ≥ 8 mg/L exhibited operon sequences identical to ChtR-P102H-*Efm*-E1162, contrasting with diverse amino-acid mutations identified in the sensor-histidine-kinase and/or in the two new transporters proteins identified in isolates with a CHX-MIC ≤ 4 mg/L and lacking ChtR-P102H.

Conclusions: The complete characterization of the ChtR-P102H-operon, highly conserved among *Efm* with high CHX-MICs, is here firstly described. Its wide distribution in *Efm* of diverse clades and sources suggests occurrence of horizontal transfer events. The role of each ChtR-operon protein in the CHX-tolerance as well as the occurrence of other CHX tolerance mechanisms in isolates with MIC ≥ 8 mg/L and lacking ChtR-P102H deserves further research.

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